NC1180 MULTISTATE PROJECT ANNUAL REPORT 2023

Tittle: Control of endemic, emerging, and re-emerging poultry respiratory diseases Chair - Maricarmen García - University of Georgia Secretary - Brian Jordan- University of Georgia Administrative Advisor- Don Reynolds – University of Nebraska

Period the Report Covers: (November 1st, 2022 – August 1^{4th}, 2023)
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Brief summary of minutes of annual meeting: It should be noted that the annual meeting was held earlier than previous years due to the impending project renewal. The meeting began with a Zoom meeting whereby, recognized experts from the poultry sectors of commercial broiler chickens, laying chickens and turkeys provided valuable information on the most critical research priorities in their respective poultry fields. This was the first time the NC1180 group has solicited stakeholder input at their annual meeting and this event proved invaluable for developing the project renewal. Following the stakeholder input meeting there was discussion on the project renewal process followed by station reports.

ACCOMPLISHMENTS:

OBJECTIVE 1 - Investigate the ecology of poultry respiratory diseases and their role in poultry flocks. Bacterial diseases- CA in collaboration with IA and MD have established a genotyping assay for Avibacterium paragallinarum (A.P.), the causative agent of infectious coryza (IC), different genotypes were established along the three main serotypes (A, B and C). Results indicated that current strains causing outbreaks of the disease in the U.S. include a genotype cluster within serotype C strains and few serotype C strains that share genotype sequences with serotype B strains. IA has conducted passive surveillance of layer flocks for A.P. using qPCR and found positive A.P. samples that on culture the bacteria presented nontypical (nt) colonies. Full genome sequence of the nt A.P. isolates indicated the absence of some virulence factors. MN studied the ecology of avian colibacillosis and found that strains of avian E. coli colonizing the gut are not representative of those found dominating clinical disease. MD gathered antimicrobial resistant (AMR) profiles and full genome sequences of Ornithobacterium rhinotracheale (ORT) isolates from commercial poultry in the U.S. This study revealed a significant rise in ORT antimicrobial-resistant, and ORT AMR profiles not necessarily correlate with genotypes. Viral diseases- CA conducted prevalence studies of avian hepatitis E virus (aHEV) and found that the spread of the virus is facilitated by the stress of production. Currently circulating aHEV genotypes include classical strains and viral genotypes not previously detected in the U.S. GA and DE have conducted active surveillance for IBV. GA IBV surveillance revealed wide differences in the level of infection and replication of the vaccine strains among vaccinated flocks and differences in protection against circulating non-vaccine origin strains in vaccinated flocks. In layer and breeder flocks' new viral genotypes were frequently found. DE reported a high number of ILTV cases from February to early July of 2023. A total of 450 positive ILTV cases were detected by PCR and 44 were confirmed by histopathology. Outbreak related

viruses were not genotyped. Other infections were reported together with ILTV such as IBV, ORT and E. coli. **SEPRL (EEAVD)** conducted sequence analysis of the North American (NA) H7 lineage viruses from wild birds and compared the HA sequences to H7 sequences from poultry viruses. Compared to wild birds H7 viruses, the H7 poultry viruses tended to contain minor genetic groups, suggesting either a biased sampling for AIVs in wild birds or a tendency of having reassortment with minor genetic groups prior to the virus's introduction to poultry.

OBJECTIVE 2- Develop new and improved diagnostic tools for poultry respiratory diseases.

AL is developing a protein histochemistry assay to determine changes in the tropism and virulence of newly emerging NDV strains. **GA** has developed new multiplex amplification and Nanopore sequencing genotyping assays for ILTV and IBV. The novel ILTV assay provided more accurate discrimination among ILTV genotypes of currently circulating viruses. The IBV new genotyping assay allowed the detection of multiple IBVs in a single sample, including IBVs of previously unknown genotype. **SEPRL (EEAD)** has developed a platform to sequence total RNA using a random sequencing approach with rapid and precise characterization of pathogens to facilitate the identification of multiple pathogens in multifactorial respiratory diseases cases. **IA** is in the process to optimize and validate a one-step Nanopore sequencing assay for the rapid identification and genetic characterization of poultry viral agents IBV, AI and virulent NDV directly from clinical samples. Utilizing a combination of plasmid and clone-specific markers **MN** has developed an avian pathogenic *Escherichia coli* (APEC) typing scheme which effectively differentiate between strains of higher and lower virulence. **IL** has developed several monoclonal antibodies that target chicken complement receptors which will facilitate the study of virus interactions with the complement system. **NE** has developed a whole blood assay for assessing innate immune response based on the level of the IL6 protein expression after LPS stimulation for rapid evaluation of vaccines, vaccine adjuvants, and vaccine programs responses.

OBJECTIVE 3– Elucidate the pathogenesis of poultry respiratory diseases. AL did a comprehensive study to show that IBV ARK-type viruses associated with outbreaks of the disease in broilers emerged through the selection of vaccine subpopulations and/or naturally occurring recombination events. Moreover, this study showed that although variant strains emerged from ARKDPI vaccination, this vaccine no longer induced effective protection against these variants. AL performed comparative whole genome analysis of specific pathogen free (SPF) chickens with high or low viral load of ARK-type virulent virus. Two genes in chicken chromosome 18 appear to be associated with IBV genome load differences. The role of these two genes will be further studied. AL is evaluating the expression of innate immune genes, the antibody responses in serum (systemic) and tears (local), and cell responses in the Harderian gland (HG) induced by vaccination with La Sota NDV vaccine. Similarly, AL is pursuing to identify the immune mechanisms behind AIV vaccines that elicit a rapid onset and broadly protective immunity. CA is studying the role of passive immunity (maternal antibodies) in preventing the development of chronic microscopic lesions in kidneys and oviducts of mature layers caused by the early exposure to IBV. Findings indicate that without adequate maternal antibodies, early IBV vaccination or challenge will cause microscopic lesions that could increase the incidence of disease throughout the life of the bird. GA is studying the expression of interferons (IFNs) and interferon stimulated genes (ISGs) induced by ILTV live vaccines and virulent to determine how viral strains with different degrees of virulence modulate IFN responses. IL has identified that the glycoprotein C (gC) gene of MDV is essential for viral transmission and that gC of a virulent MDV strain can compensate for gC function on MD vaccine. Swapping MDV vaccine gC for MD gC may enhance MDV-specific immunity without affecting vaccine replication and spread. IL obtained expression levels of MDV genes at the transcript and protein levels in the skin of infected chickens. Future work will resolve with genes are relevant for MDV transmission. **MD** studied the effect that IBDV infection plays in the adaptation and shedding of a mallard AI virus when inoculated into groups of chickens infected and non-infected with IBDV. IBDV exposure prolonged the shedding of the mallard AIV in chickens and increased the population diversity of the mallard AIV but did not contribute to the virus adaptation to the chicken host. SEPRL (EEAVD) conducted a study to determine the potential role of mallards as efficient reservoirs to amplify and disseminate the H5N1 clade 2.3.4.4b highly pathogenic avian influenza viruses (HPAIVs). Infection appeared to be subclinical in most ducks, but high virus infectivity and transmission were observed which highlights the potential role of mallards as efficient reservoirs to amplify and disseminate H5N1 viruses. SEPRL (ENAVD) had identified novel ILTV transcripts encoding viral immuno-modulators. The plan is to generate

knock-out mutants which lacks these homologous viral proteins and to study their pathogenesis in chickens.

OBJECTIVE 4- Develop new prevention and control strategies for poultry respiratory diseases. <u>Viral Vaccines</u>-AL developed a vaccination strategy for IBV were a recombinant NDV La Sota co-expressing the trimeric spike ectodomain (SE) of the ARK-DPI and the chicken granulocyte-macrophage colony-stimulating factor (GMCSF) (rLS/Ark.Se.GMCSF) was co-administered with the Massachusetts (Mass) live vaccine. The vaccine combination had cross-protective ability against heterologous challenge. SEPRL (ENAVD) are evaluating three vaccine platforms for in ovo delivery. One platform utilizes NDV La Sota strain as a vector. This construct can be safely administered in ovo and it was shown to induced significant protection against MDV when expressing MDV gB. The second vaccine platform utilize MDV serotype II strain as a vector to share expression of multiple antigens when co-administered with HVT. The third vaccine platform are mRNA-based vaccines expressing immunogens of ILTV and IBV. SEPRL (EEAVD) tested the protection efficacy of four available AIV H5N1 vaccines, three inactivated and one an RNA particle vaccine, against highly pathogenic Clade 2.3.4.4b virus in chickens. This study confirmed the efficacy of currently available H5N1 vaccines and assessed the potential application of serologic test to differentiate infected (after vaccination) from non-infected or vaccinated animals. SEPRL (EEAVD) tested the adjuvant potential of ODN-1826 and Imiquimod, agonist of pathogen recognition receptors, to increase efficacy of Newcastle disease virus vaccines. Vaccinated + adjuvant groups of chickens had higher and more uniform antibody titers and challenge virus shedding was reduced indicating that agonist of pathogen recognition receptors are effective mucosal vaccine enhancers. Bacterial Vaccines- CA assessed the efficacy of a commercially available trivalent inactivated vaccine against infectious coryza (IC) that contains three A.P. serovars. Chickens were protected against clinical signs and reduction of bacteria shedding was detected. Treatments- CA studied the protective effects of IBV passive immunity by investigating various methods to administered (spray, in-ovo injection) IBV hyperimmune serum to day-of-age chickens. Administration of antibodies via spray at day of age was effective in reducing clinical signs and viral shedding post challenge. This treatment may prevent long-term reproductive effects of early IBV infection. Biosecurity-Education-Outreach programs. NE continue the use of the "Big Red Biosecurity Program" outreach efforts provided information on how to improve biosecurity to avoid the introduction of Avian Influenza. MD has established an extension program to facilitate passive and active control of HPAI outbreaks. The group perform biosecurity compliance audits and implement risk-based planning to improve outbreak responses.

IMPACTS: Understanding the epidemiology of respiratory diseases in the U.S., through surveillance, mapping, and genetic characterization strategies is crucial to establish successful prevention and control strategies including vaccination, disease management, and biosecurity. The research accomplished in this project has allowed laboratories across the U.S. to implement new approaches to detect and type bacterial and viral pathogens affecting poultry and to establish novel approaches to study immune responses to these pathogens. These new assays are streamlining diagnostics procedures and facilitating research by providing a better understanding of the host-pathogen interaction. Understanding how vaccines allow emergence of new variants, how maternal antibodies can ameliorate chronic disease, how immunosuppressive diseases of poultry amplify H5N1 HPAI infections, and the identification of viral genes that contribute to viral transmission or immune evasion are mayor contributions of this project in the control of respiratory diseases of poultry. Knowledge on the nature of early antiviral and adaptive protective immune responses induced by vaccination is fundamental for the future design of vaccines against these diseases. Prevention and control of respiratory diseases of poultry require simultaneous strategies including robust biosecurity, effective education programs, development of safer more effective novel vaccines and optimal use of current vaccines. The NC1180 project contributes with innovative solutions in these areas that have a strong impact in the health and production of poultry.

PUBLICATIONS

** Names in bold denote collaborations between members of the NC1180 group.

Becerra, R., Maekawa, D., & García, M. (2023). Protection Efficacy of Recombinant HVT-ND-LT and the Live Attenuated Tissue Culture Origin Vaccines Against Infectious Laryngotracheitis Virus When Administered Individually or in Combination. Avian diseases, 67(2), 145–152. https://doi.org/10.1637/aviandiseases-D-23-00010

Botchway, P. K., Amuzu-Aweh, E. N., Naazie, A., Aning, G. K., Otsyina, H. R., Saelao, P., Wang, Y., Zhou, H., Walugembe, M., Dekkers, J., Lamont, S. J., Gallardo, R. A., Kelly, T. R., Bunn, D., & Kayang, B. B. (2022). Host response to successive challenges with lentogenic and velogenic Newcastle disease virus in local chickens of Ghana. Poultry science, 101(11), 102138. https://doi.org/10.1016/j.psj.2022.102138

Bueno, I., Ricke, I., Hwang, H., Smith, E., Nault, A., Johnson, T. J., & Singer, R. S. (2023). Efficacy of Antibiotic and Non-antibiotic Interventions in Preventing and Treating Necrotic Enteritis in Broiler Chickens: A Systematic Review. Avian diseases, 67(1), 20–32. https://doi.org/10.1637/aviandiseases-D-22-00069

Buter, R., Feberwee, A., de Wit, S., Heuvelink, A., da Silva, A., Gallardo, R., Soriano Vargas, E., Swanepoel, S., Jung, A., Tödte, M., & Dijkman, R. (2023). Molecular characterization of the HMTp210 gene of Avibacterium paragallinarum and the proposition of a new genotyping method as alternative for classical serotyping. Avian pathology : journal of the W.V.P.A, 52(5), 362–376. https://doi.org/10.1080/03079457.2023.2239178

Butt, S. L., Kariithi, H. M., Volkening, J. D., Taylor, T. L., Leyson, C., Pantin-Jackwood, M., Suarez, D. L., Stanton, J. B., & Afonso, C. L. (2022). Comparable outcomes from long and short read random sequencing of total RNA for detection of pathogens in chicken respiratory samples. Frontiers in veterinary science, 9, 1073919. <u>https://doi.org/10.3389/fvets.2022.1073919</u>

Campler, M. R., Hashish, A., Ghanem, M., El-Gazzar, M. M., & Arruda, A. G. (2023). Space-Time Patterns of Poultry Pathogens in the USA: A Case Study of Ornithobacterium rhinotracheale and Pasteurella multocida in Turkey Populations. Pathogens (Basel, Switzerland), 12(8), 1004. https://doi.org/10.3390/pathogens12081004

Chaney, E., Miller, E. A., Firman, J., Binnebose, A., Kuttappan, V., & Johnson, T. J. (2023). Effects of a postbiotic, with and without a saponin-based product, on turkey performance. Poultry science, 102(5), 102607. <u>https://doi.org/10.1016/j.psj.2023.102607</u>

Clark, A. A., Eid, S., Hassan, M. K., Carter, K., & Swayne, D. E. (2022). Reducing zoonotic avian influenza transmission at household poultry slaughter using a behaviour change tool for limited literacy audiences. Zoonoses and public health, 69(8), 956–965. <u>https://doi.org/10.1111/zph.12993</u>

Criado, M. F., Kassa, A., Bertran, K., Kwon, J. H., Sá E Silva, M., Killmaster, L., Ross, T. M., Mebatsion, T., & Swayne, D. E. (2023). Efficacy of multivalent recombinant herpesvirus of turkey vaccines against high pathogenicity avian influenza, infectious bursal disease, and Newcastle disease viruses. Vaccine, 41(18), 2893–2904. <u>https://doi.org/10.1016/j.vaccine.2023.03.055</u>

Delago, J., Miller, E. A., Flores-Figueroa, C., Munoz-Aguayo, J., Cardona, C., Smith, A. H., & Johnson, T. J. (2023). Survey of clinical and commensal Escherichia coli from commercial broilers and turkeys, with emphasis on high-risk clones using APECTyper. Poultry science, 102(7), 102712. https://doi.org/10.1016/j.psj.2023.102712

Espejo, R., Breedlove, C., da Silva, L. F., Joiner, K., Toro, H. (2023). Cross-Protection Conferred by Combined Vaccine Containing Infectious Bronchitis Virus Attenuated Massachusetts and Recombinant LaSota Virus Expressing Arkansas Spike. Avian diseases. *Submitted*.

Ghanem, M., Hashish, A., Chundru, D., & El-Gazzar, M. (2023). Complete Genome Sequence and Annotation of Malacoplasma iowae Type Strain 695, Generated Using PacBio Sequencing. Microbiology resource announcements, 12(1), e0049022. <u>https://doi.org/10.1128/mra.00490-22</u>

Ghorbani, A., Ngunjiri, J. M., Edward C Abundo, M., Pantin-Jackwood, M., Kenney, S. P., & Lee,

C. W. (2023). Development of in ovo-compatible NS1-truncated live attenuated influenza vaccines by modulation of hemagglutinin cleavage and polymerase acidic X frameshifting sites. Vaccine, 41(11), 1848–1858. <u>https://doi.org/10.1016/j.vaccine.2023.01.018</u>

Hashish, A., Johnson, T. J., Chundru, D., Williams, M. L., Sato, Y., Macedo, N. R., Clessin, A., Gantelet, H., Bost, C., Tornos, J., Gamble, A., LeCount, K. J., Ghanem, M., Boulinier, T., & El-Gazzar, M. (2023). Complete Genome Sequences of Two Pasteurella multocida Isolates from Seabirds. Microbiology resource announcements, 12(4), e0136522. https://doi.org/10.1128/mra.01365-22

Hashish, A., Johnson, T. J., Smith, E., Chundru, D., Williams, M. L., Macedo, N. R., Sato, Y., Ghanem, M., & El-Gazzar, M. (2023). Complete Genome Sequences of Three Ornithobacterium rhinotracheale Strains from Avian Sources, Using Hybrid Nanopore-Illumina Assembly. Microbiology resource announcements, 12(2), e0105922. https://doi.org/10.1128/mra.01059-22

Helmy, Y. A., El-Adawy, H., Sanad, Y. M., & Ghanem, M. (2023). Editorial: Food safety and public health. Frontiers in microbiology, 14, 1169139. <u>https://doi.org/10.3389/fmicb.2023.1169139</u>

Johnson, T. J., Miller, E. A., Flores-Figueroa, C., Munoz-Aguayo, J., Cardona, C., Fransen, K., Lighty, M., Gonder, E., Nezworski, J., Haag, A., Behl, M., Kromm, M., Wileman, B., Studniski, M., & Singer, R. S. (2022). Refining the definition of the avian pathogenic Escherichia coli (APEC) pathotype through inclusion of high-risk clonal groups. Poultry science, 101(10), 102009. https://doi.org/10.1016/j.psj.2022.102009

Jude, R., da Silva, A. P., Rejmanek, D., Shivaprasad, H. L., Stoute, S., Jerry, C., Gallardo, R. A. (2023). Whole genome sequence of a novel genotype VIII infectious bronchitis virus isolated from California layers in 2021. ASM Microbiology Resource Announcements. *Submitted*.

Jude, R., Jordan, B., Muller-Slay A., Luciano, R., da Silva, A.P., Gallardo, R. A. (2023). Mitigation of False Layer Syndrome Through Maternal Antibodies Against Infectious Bronchitis Virus. *Submitted*.

Kariithi, H. M., Christy, N., Decanini, E. L., Lemiere, S., Volkening, J. D., Afonso, C. L., & Suarez, D. L. (2022). Detection and Genome Sequence Analysis of Avian Metapneumovirus Subtype A Viruses Circulating in Commercial Chicken Flocks in Mexico. Veterinary sciences, 9(10), 579. https://doi.org/10.3390/vetsci9100579

Kariithi, H. M., Suarez, D. L., Davis, J. F., Dufour-Zavala, L., Olivier, T. L., Williams-Coplin, D., Bakre, A., & Lee, C. W. (2023). Genome Sequencing and Characterization of an Avian Orthoavulavirus 1 VG/GA-like Isolate with a Unique Fusion Cleavage Site Motif. Avian diseases, 67(1), 33–41. <u>https://doi.org/10.1637/aviandiseases-D-22-00064</u>

Kwon, J. H., Bertran, K., Lee, D. H., Criado, M. F., Killmaster, L., Pantin-Jackwood, M. J., & Swayne, D. E. (2023). Diverse infectivity, transmissibility, and pathobiology of clade 2.3.4.4 H5Nx highly pathogenic avian influenza viruses in chickens. Emerging microbes & infections, 12(1), 2218945. https://doi.org/10.1080/22221751.2023.2218945

Lee K., Breedlove, C., Khalid, Z., Sheikhsamani, E., Alrubaye, A., Dridi, S., Pummill, J., Toro H., Rhoads, D. D. (2023). Whole genome resequencing suggests limited genetic basis for wide variation in infectious bronchitis viral load in challenged naïve white leghorn chickens. *Submitted*.

Lee, C. W., Kc, M., Ngunjiri, J. M., Ghorbani, A., & Lee, K. (2023). TLR3 and MDA5 Knockout DF-1 cells Enhance Replication of Avian Orthoavulavirus 1. Avian diseases, 67(1), 94–101. https://doi.org/10.1637/aviandiseases-D-22-00065

McDuie, F., Overton, C. T., Lorenz, A. A., Matchett, E. L., Mott A. L., Mackell, D. A., Ackerman J. T., de la Cruz S. E. W., Patil, V. P., Prosser, D. J., Takekawa, J. Y., Orthmeyer, D. L., Pitesky, M. E., Díaz-Muñoz, S. L., Riggs, B. M., Gendreau, J., Reed, E. T., Petrie, M. J., Williams, C. K, Buller J.J., Hardy, M. J., Ladman, B. S., Legagneux, P., Bêty, J., Thomas, P. J., Rodriguez, J., Lefebvre, J.,

Casazza, M. L. (2023). Predicting H5N1 avian influenza spread with an empirical model of bird movement. Ecology Letters-*Submitted*

Ponnuraj, N., Akbar, H., Arrington, J. V., Spatz, S. J., Nagarajan, B., Desai, U. R., & Jarosinski, K. W. (2023). The alphaherpesvirus conserved pUS10 is important for natural infection and its expression is regulated by the conserved Herpesviridae protein kinase (CHPK). PLoS pathogens, 19(2), e1010959. <u>https://doi.org/10.1371/journal.ppat.1010959</u>

Ramsubeik, S., Stoute, S., Gallardo, R. A., Crossley, B., Rejmanek, D., Jude, R., & Jerry, C. (2023). Infectious Bronchitis Virus California Variant CA1737 Isolated from a Commercial Layer Flock with Cystic Oviducts and Poor External Egg Quality. Avian diseases, 67(2), 212–218. https://doi.org/10.1637/aviandiseases-D-23-00014

Smith, J., Alfieri, J. M., Anthony, N., Arensburger, P., Athrey, G. N., Balacco, J., Balic, A., Bardou, P., Barela, P., Bigot, Y., Blackmon, H., Borodin, P. M., Carroll, R., Casono, M. C., Charles, M., Cheng, H., Chiodi, M., Cigan, L., Coghill, L. M., Crooijmans, R., Zhou, H. (2022). Fourth Report on Chicken Genes and Chromosomes 2022. Cytogenetic and genome research, 162(8-9), 405–528. https://doi.org/10.1159/000529376

Spackman, E., Pantin-Jackwood, M. J., Lee, S. A., & Prosser, D. (2023). The pathogenesis of a 2022 North American highly pathogenic clade 2.3.4.4b H5N1 avian influenza virus in mallards (Anas platyrhynchos). Avian pathology : journal of the W.V.P.A, 52(3), 219–228. https://doi.org/10.1080/03079457.2023.2196258

Spatz, S., García, M., Fuchs, W., Loncoman, C., Volkening, J., Ross, T., Riblet, S., Kim, T., Likens, N., & Mettenleiter, T. (2023). Reconstitution and Mutagenesis of Avian Infectious Laryngotracheitis Virus from Cosmid and Yeast Centromeric Plasmid Clones. Journal of virology, 97(4), e0140622. <u>https://doi.org/10.1128/jvi.01406-22</u>

Tsaxra, J. B., Gallardo R. A., Abolnik, C., Chengula, A., Msoffe, P. M., Muhairwa, A. P., Phiri, T., Mushi, J. R., Chouicha, N., Mollel, E. L., Zhou, H., Kelly, T. R. (2023). Spatio-temporal Patterns and Prevalence of Newcastle Disease Virus at Mawenzi Live Bird Market in Morogoro Municipality, Tanzania. Transboundary and emerging diseases. *Submitted*.

Tsaxra, J. B., Gallardo R. A., Abolnik, C., Jude, R., Chengula, A., Msoffe, P. M., Muhairwa, A. P., Phiri, T., Mushi, J. R., Chouicha, N., Mollel, E. L., Zhou, H., Kelly, T. R. (2023). Spatio-temporal Patterns and Prevalence of Newcastle Disease Virus at Mawenzi Live Bird Market in Morogoro Municipality, Tanzania. Tropical animal health and production. *Submitted*.

Tudeka, C. K., Aning, G. K., Naazie, A., Botchway, P. K., Amuzu-Aweh, E. N., Agbenyegah, G. K., Enyetornye, B., Fiadzomor, D., Saelao, P., Wang, Y., Kelly, T. R., Gallardo, R., Dekkers, J. C. M., Lamont, S. J., Zhou, H., & Kayang, B. B. (2022). Response of three local chicken ecotypes of Ghana to lentogenic and velogenic Newcastle disease virus challenge. Tropical animal health and production, 54(2), 134. <u>https://doi.org/10.1007/s11250-022-03124-8</u>

Volkening, J. D., Spatz, S. J., Ponnuraj, N., Akbar, H., Arrington, J. V., Vega-Rodriguez, W., & Jarosinski, K. W. (2023). Viral proteogenomic and expression profiling during productive replication of a skin-tropic herpesvirus in the natural host. PLoS pathogens, 19(6), e1011204. https://doi.org/10.1371/journal.ppat.1011204

Walugembe, M., Naazie, A., Mushi, J. R., Akwoviah, G. A., Mollel, E., Mang'enya, J. A., Wang, Y., Chouicha, N., Kelly, T., Msoffe, P. L. M., Otsyina, H. R., Gallardo, R. A., Lamont, S. J., Muhairwa, A. P., Kayang, B. B., Zhou, H., & Dekkers, J. C. M. (2022). Genetic Analyses of Response of Local Ghanaian Tanzanian Chicken Ecotypes to a Natural Challenge with Velogenic Newcastle Disease Virus. Animals: an open access journal from MDPI, 12(20), 2755. https://doi.org/10.3390/ani12202755

Youk, S., Leyson, C., Killian, M. L., Torchetti, M. K., Lee, D. H., Suarez, D. L., & Pantin-Jackwood, M. J. (2022). Evolution of the North American Lineage H7 Avian Influenza Viruses in Association

with H7 Virus's Introduction to Poultry. Journal of virology, 96(14), e0027822. https://doi.org/10.1128/jvi.00278-22