

#### **Table Of Contents**

Statement Of Issues And Justification	1
Prerequisite Criteria: Mission	2
Prerequisite Criteria: How does this NRSP pertain to a national issue	3
The advantage of doing this work as a multistate project	6
What the likely impacts will be from successfully completing the work	
Rationale: Priority Establish by ESCOP/ESS	
Rationale: Relevance to Stakeholders Stakeholder needs Measuring stakeholder use	8 10
<ul> <li>Objectives and Projected Outcomes <ol> <li>Identify the animal drug needs for minor species and minor uses</li> <li>major species.</li> <li>Generate and disseminate data for the safe, effective and legal use of drugs used primarily in therapy or reproductive management of minor animal species.</li> <li>Facilitate FDA/ CVM approvals of drugs for minor species and minor uses</li> <li>Provide alternatives to antibiotic use in food animals through the identification and FDA/CVM approval of naturally occurring biotherapeutics and feed additives.</li> </ol> </li> </ul>	11 12 13
5. Expected Outcomes	13 14 14
Management, Budget and Business Plan         1. Organizational structure         2. Funding activities         3. Research         3.1. Research planned         4. FDA/CVM approval         5. Assessment of outcomes         5.1. Productivity         5.2. Completion of original objectives	16 16 18 19 19 19 19
Integration	20
Outreach, Communication and Assessment	20
Projected Participation	22
Table 1. Economic Impact Of Minor Animal Species By State As Of 2013         Table 2. Animal Drug Approvals And Current NRSP-7 Activity By Species         Table 3. Minor Use Animal Drug Program Total Funding 2009-2013 By Source         Figure 1. Flow Chart Outlining The Process For Selection Of Drugs For Testing In The NRSP-7         Minor Use Animal Drug Program	.24 25
Peer-Reviewed Publications	
Literature Cited	
External Links	39
Appendix E Part 1 Participant List; Part 2 Research Summary; Part 3 Extension Summary Appendix F NRSP-7 Budget Request Summary – 4 pages.	

### NRSP007: A National Agricultural Program for Minor Use Animal Drugs

Duration:	October 1, 2015 to September 30, 2020
Administrative Advisor(s):	[Margaret E. Smith (main) NE] [Philip Elzer S]
	[Frank D. Galey W] [George Smith NC]
NIFA Rep:	[Gary Sherman]

#### **Statement Of Issues And Justification**

Over the last two previous decades, the lack of approved drugs for minor species had placed US producers at a competitive disadvantage. Globalization of food markets allows countries with more products available for such uses to produce more animals and to export food products to the US. This has proved especially hard for the sheep, farmed fish and shrimp, and honey industries.

More recently, government agencies around the world and the public have become concerned about the continued use of antibiotics in food producing animals. Internationally, scientists have provided strong evidence that antibiotic use in food-producing animals can have a negative impact on public health through the following sequence of events: (1) Use of antibiotics in food-producing animals allows antibiotic-resistant bacteria to thrive while susceptible bacteria are suppressed or die; (2) Resistant bacteria can be transmitted from food-producing animals to humans through the food supply; (3) Resistant bacteria can cause infections in humans; and (4) Infections caused by resistant bacteria can result in adverse human health consequences. Because of the link between antibiotic use in food-producing animals and the occurrence of antibiotic-resistant infections in humans, the Food and Drug Administration/Center for Veterinary Medicine (FDA/CVM) and Centers for Disease Control (CDC) encourage continuing efforts to minimize inappropriate use of antibiotics in humans and animals.

NRSP-7 is an efficient and important program that supports research specifically to gain FDA/CVM approval of new animal drugs for use in minor animal species of agricultural importance. These minor species include sheep, goats, farmed deer, all cultured fish and crustaceans, game birds such as pheasants, partridges, and quail, emus, and honey bees. Data supplied by the Program provided for the publication of 52 New Animal Drug Applications and modification of 73 label claims to include minor species, an overall average of 1.6 New Animal Drug Applications per year and 2.3 new label claims per year. Included in these Public Master Files were one for rabbits, nine for game birds, 16 for fish, lobster and shrimp, 15 for meat and dairy goats, eight for bison and reindeer, one for foxes and two for honey bees. For the 5-year period of this review, NSRP-7 was responsible for three Public Master Files; they included Progesterone Solid Matrix for sheep (NADA 141-302), Lincomix for the control of American foulbrood in honey bees (NADA 111-636) and Chloramine-T for the control of mortality in freshwater-reared salmonids due to bacterial gill diseases (NADA 141-423).

Additionally, NRSP-7 has published 211 articles in peer-reviewed journals, averaging 6.6 per year over the term of the program. For the last five years, however, publications have increased to nearly 10 per year. Thus, although FDA/CVM drug approvals have waned due to increasing costs, the Program has increased its efforts to supply critical data needs to minor species producers. The data generated by the Program is also shared with the Food Animal Residue Avoidance Database (FARAD program to further increase visibility.

While an average of slightly more than one drug approval per year does not begin to address the needs of all these species, it is a remarkable performance for such a small and minimally funded program. The Animal Health Institute, which represents veterinary pharmaceutical companies, states that bringing a major new animal drug to market with FDA/CVM approval

generally takes 7 to 10 years and costs up to \$100 million (<u>http://www.ahi.org/about-animl-medicines/industry-statistics/</u>). While a minor species use is generally supplemental to a major species approval, it takes nearly as long to complete the requirements and generally still costs several million dollars. NRSP-7 has leveraged partnerships with academia and with producer groups to stretch their budget, but that has become more and more difficult as costs have risen and funding has been reduced.

Even with the estimated increased cost per drug approval in recent years, the NRSP-7 program continues to demonstrate remarkable efficiency and cost effectiveness. Compared to an average investment of the pharmaceutical industry estimated at \$10 to \$25 million in 2015 for adding a label claim to an existing veterinary drug, information generated for additional label claims by the NRPS-7 program costs only approximately 15 to 35% of pharmaceutical industry costs (http://www.ahi.org/about-animal-medicines/industry-statistics/).

The NRSP-7 is the only national program designed and organized to address the issues of the prudent use of antibiotics, anthelmintics and production drugs in minor species of food- and fiber-producing animals. For 27 years, the Program was funded through a USDA special research grant administered by NIFAin cooperation with the NRSP-7 Technical Committee. Currently, however, NRSP-7 has been dependent on "off-the-top" Regional Research funds allocated to the Minor Use Program. Support for NRSP-7 also comes from pharmaceutical companies, producers and universities in the form of "in kind" contributions for Regional Coordinators. The program also receives significant "in-kind support from several other sources including the institutions conducting the research (state agriculture experiment stations, colleges of veterinary medicine, federal laboratories), animal producer groups through contributions of animals for research, and pharmaceutical companies. For example, /CVM cash and in-kind support was \$1.8 million or 1.9-times Hatch funding received during the previous 5-year period of 2009 to 2014. For this same period, support by Stakeholders (producers and pharmaceutical companies) was \$5.7 million or 6-times Hatch funding. All outside support totaled \$9.7 million or 9.2-times the Hatch funding received by the MUADP over these five years.

Over the next five years, the Program will pursue additional funding support from the Minor Use Minor Species Program of the FDA/CVM and developers of organic or natural alternatives to antimicrobial and pesticide use in food animals. With its recent inclusion in the 2014 Farm Bill, the Program has greater leverage with Congress and NIFA to resume financial support of its mandated research objectives. Stakeholders will also be engaged to further Congressional interest in tighter control of antimicrobial use in food animals through NRSP-7 research. These efforts, we believe, will assist the Program to be federally funded at a once again at more adequate amounts.

#### Prerequisite Criteria: Mission

The activity of a National Research Support Project (NRSP) focuses on the development of enabling technologies, support activities (e.g., collect, assemble, store, and distribute materials, resources and information), or the sharing of facilities needed to accomplish high priority research, but which is not of itself primarily research. The overall objectives of NRSP-7 the USDA Minor Use Animal Drug Program are to provide for human food safety and animal health through the judicious use of antibiotics and therapeutics in food and fiber producing animals. To accomplish these goals, the Program has developed the following missions:

- 1. Identify animal drug needs for minor species and minor uses in major species,
- 2. Generate and disseminate data for safe and effective therapeutic applications,
- Facilitate FDA/CVM approvals for drugs identified as a priority for a minor species or minor use, and

4. Provide alternatives to antibiotic use in food animals through the identification and FDA/CVM approval of naturally occurring biotherapeutics and feed additives.

Minor uses include minor species (all species except dogs, cats, horses, cattle, swine, chickens, and turkeys), while minor uses in major species are those that occur infrequently or in limited geographical locations. The primary emphasis of the Program is on food-and/or fiber- (hair, wool, fur, feathers or hide) producing minor species with a secondary interest in non-food animals such as bees and tropical fish.

With this review, the Program is expanding its mission to providing natural alternatives to antibiotic use in food animals (#4 above). This expansion has been considered necessary to respond to the critical national issue of antimicrobial resistance relating to the use of antibiotics in animals feeds as growth promoters. This aspect of the Program has been modeled after the highly successful biopesticides initiative of NRSP-4 (IR-4).

To accomplish these goals, NRSP-7 functions through the coordination of efforts among animal producers, pharmaceutical manufacturers, Food and Drug Administration/Center for Veterinary Medicine, United States Department of Agriculture/Cooperative State Research, Education, and Extension Service, universities, State Agricultural Experiment Stations and veterinary medical colleges throughout the country.

### Prerequisite Criteria: How does this NRSP pertain to a national issue?

In February of 2014 Congress and the President recognized the national role of NRSP-7 in H. R. 2642 the AGRICULTURAL ACT OF 2014, known as the 2014 Farm Bill. **SEC. 7404**. **COMPTETITIVE, SPECIAL, AND FACILITIES RESERCH GRANT ACT** includes among its (b) **PRIORITY AREAS** (1)(C) "(x) the identification of animal drug needs and the generation and dissemination of data for safe and effective therapeutic application of animal drugs for minor uses and minor uses of such drugs in major species." Further, Congressional support for NRSP-7 was provided by the Managers on the part of the House and the Senate in the **JOINT EXPLANATORY STATEMENT OF THE COMMITTEE OF CONFERENCE.** Page 166 of this document establishes the NRSP-7 Program with the wording, "This section … establishes National Research Support Project-7 for research on drugs for use in minor animal species. (Section 7307)." On page 167 the JOINT STAGEMENT notes that, "The Managers encourage the Director of NIFA to continue to support National Research Support Project-7 and to work cooperatively with the Center for Veterinary Medicine of the Food and Drug Administration to facilitate the development and approval of drugs for minor species and minor uses for major species. (Section 7406)."

Additionally, globalization of food markets has allowed countries with greater animal drug availability an advantage when competing with our sheep, goat, rabbits, deer, farmed shrimp and fish, and honey industries. One-third of the lamb and 82% of venison consumed in the US comes from Australia and New Zealand. Nearly 90% of the commercially farmed shrimp are imported. Additionally, two-thirds of the honey consumed in the US is imported and half of that honey comes from China. In order to complete with these countries, American producers are forced to use therapeutics not approved in minor species.

Government agencies around the world and the public have become concerned about the continued use of antibiotics in food producing animals. Scientists in the U.S. and Europe have provided strong evidence that antibiotic use in food-producing animals can have a negative impact on public health. Antibiotics must be used judiciously in humans and animals because both uses contribute to the emergence, persistence, and spread of resistant bacteria. Resistant bacteria in food-producing animals are of particular concern. Food animals serve as a reservoir

of resistant pathogens and resistance mechanisms that can directly or indirectly result in antibiotic resistant infections in humans. Some bacteria have become resistant to more than one type of antibiotic, which makes it more difficult to treat the infections they cause. Preserving the effectiveness of antibiotic drugs is vital to protecting human and animal health.

The economic impact of minor animal species agriculture in the United States is great, but at risk. The United States gross annual farm gate income from production of specialty animal species has been estimated by producer groups at over \$4.8 billion. Further, these farm gate revenues produce an economic stimulus to the US Gross Domestic Product estimated at another \$37 billion. Table 1 provides a breakdown of theses national figures by state. Lack of approved drugs for these producers is seriously threatening the growth and long-term viability of these collective industries and the security of our food supply. While the cumulative contribution of minor species to agricultural income is great, the return to pharmaceutical companies for research on therapeutics for this category, by species is small and generally unprofitable.

Because of this substantial investment in time and resources, pharmaceutical companies must be assured that the drug will have a reasonable potential for profit. Therefore most drug approvals are sought only for those animal species that are produced in sufficient numbers to support large volume sales, specifically cattle, swine, chickens and turkeys. There is little economic incentive for pharmaceutical firms to generate data necessary to seek FDA/CVM approval of drugs in minor species; hence, very few drugs are available for management of diseases in these minor species. Inequities in drug availability represent serious management and economic problems for producers for minor species. Today, more than half of all commercially led pharmaceutical R&D in the veterinary medical field is focused on developing products for companion animals, and the emphasis on this sector is likely to increase in coming years, as companion animals live longer, and more diseases of old age are diagnosed and treated<sup>2</sup>.

The FDA/CVM has been aware that veterinarians and livestock producers were using unapproved drugs for minor species without the safeguards that approved drugs carry. Additionally, little peer-reviewed literature existed to provide veterinarians with sufficient information for rationale extra-label use. Such unapproved drug use could not only cause detrimental effects to the animals being treated, but could also lead to the persistence of drug residues in animal products intended for human consumption. Efforts were and continue to be necessary to provide US animal producers with safe and effective means to compete in a global market, while assuring US consumers a safe and wholesome food supply.

The FDA has taken the position to promote the judicious use of antibiotics that are important in treating humans. This strategy recommends that such antibiotics should be used in food-producing animals only under veterinary oversight and only to address animal health needs, not to promote growth. The American Veterinary Medical Association has, in concert, developed a position statement on the judicious therapeutic use of antimicrobials, essentially limiting any use to under the direct supervision of a veterinarian.

*Background* - In 1976, the FDA/CVM initiated an extensive study of the minor use of animal drugs through the efforts of a minor use/minor species drug committee. This committee, comprised representatives of the FDA's then Bureau of Veterinary Medicine and Bureau of Foods, the U.S. Department of Agriculture (USDA), the pharmaceutical industry, and animal producer groups. They identified the scope of the problem as a lack of approved drugs for (1) diseases of minor species and (2) the principal minor diseases of major species. The committee identified the principal diseases for which drugs were not available in the minor species. In summary, private sponsors had supported approvals for the use of minor use drugs as follows:

none for rabbits, one for ducks and pheasants (none for other game birds), two for food fish, four for goats and 21 for sheep. Minor and specialty use needs have continued to accumulate, leaving the producers of these species without the drugs necessary for disease prevention and control. A definite need has been established for approval of minor use veterinary drugs and the scope of the problem was defined. This need was also affirmed by various grower organizations.

Additionally, the committee recognized that the livestock industry in the United States relies heavily on the judicious use of drugs for the treatment of diseases in food animals. Without these drugs, animal suffering and mortality would greatly increase, as would the cost of producing animal-derived food products. However, before a drug can be marketed for use in a food animal species, it must be shown to be safe to the human consumer of the animal-derived food, and safe and efficacious in the target animal.

The process of generating the safety and efficacy data necessary for FDA/CVM approval of a drug is costly and time-consuming. In 1999, it was estimated that the cost to a pharmaceutical company for research necessary to obtain FDA/CVM approval for a new drug exceeded \$20 million, and required 8 to 10 years of concentrated research effort <sup>1</sup>. More recently, issues relating to (1) escalating costs in the development of analytical methods, (2) concerns over antimicrobial resistance in human medicine, and (3) increased environmental testing have increased veterinary drug approval costs dramatically<sup>2</sup>. Drug approvals are generally species and disease specific and additional label claims also come with considerable added expense. Pharmaceutical company estimates place the cost of simply adding a label claim to an FDA/CVM approved drug at \$10 to \$25 million<sup>3, 4</sup>.

In 1982, the IR-4 Animal Drug Program was established as part of the overall IR-4 Minor Use Pesticide Management Program. Since that time the animal portion has been established as a national means of securing approved drugs and as a conduit between the animal industries and the FDA/CVM.

In December 1990, the USDA/CSRS requested a peer review of the IR-4 program, including both the pesticide portion and the minor use animal component. A reorganization of the minor use animal drug section was one of the recommendations of the Review Team. This Change was carried out with the development of a separate Minor Use Animal Drug Technical Committee that reported to the IR-4 Administrative Advisors.

In 1992, IR-4 Administrative Advisors recommended that with the change from interregional Projects (IRs) to National Research Support Projects (NRSPs), as well as the experience gained under the reorganized IR-4 Project, that the two programs (pesticide and animal) be separated into two projects. In 1993, NRSP-7 was thus created as the Minor Use Animal Drug Program.

Congress has considered bills to promote drug availability for minor species and for minor uses in major species. The Animal Medicinal Drug Uses Clarification Act of 1994 [AMDUCA] and the Animal Drug Approval Act [ADAA] have expanded "extra label" uses for minor species. The limitations imposed by AMDUCA on extra-label drug use in feeds, however, proved to be a major problem to aquaculture and game bird industries and a guidance document has outlined conditions where limited extra-label use of approved formulations will be permitted under conditions of a valid veterinarian-client-patient relationship. The Minor Use Animal Drug Program is the only organized State/Federal effort to address the inadequate number of FDA/CVM approved drugs available for minor-use species and has been responsible for nearly all of the progress made in the approval of minor-use/minor-species drugs. It also hopes to become the most experienced program to address the excessive use of antibiotics as feed additives. Most recently, in February of 2014 Congress and the President recognized the national role NRSP-7 in H. R. 2642 the AGRICULTURAL ACT OF 2014, known as the 2014 Farm Bill. SEC. 7404. COMPTETITIVE, SPECIAL, AND FACILITIES RESEARCH GRANT ACT includes among its (b) PRIORITY AREAS (1)(C) "(x) the identification of animal drug needs and the generation and dissemination of data for safe and effective therapeutic application of animal drugs for minor uses and minor uses of such drugs in major species." Further Congressional support for NRSP-7 was provided by the Managers on the part of the House and the Senate in the JOINT EXPLANATORY STATEMENT OF THE COMMITTEE OF CONFERENCE. Page 166 of this document establishes the NRSP-7 Program with the wording, "This section … establishes National Research Support Project-7 for research on drugs for use in minor animal species. (Section 7307)." On page 167 the JOINT STAGEMENT notes that, "The Managers encourage the Director of NIFA to continue to support National Research Support Project-7 and to work cooperatively with the Center for Veterinary Medicine of the Food and Drug Administration to facilitate the development and approval of drugs for minor species and minor uses for major species. (Section 7406)."

**The advantages of doing the work as a multistate project** - There are two critical advantages for organizing the Minor Use Animal Drug Program as a multistate project. First, farm production of minor species represents a highly fragmented industry. Of the eight or so farming industries that focus on the production of minor species, none represent more than ten states in the US. To cite only two examples, catfish farming is concentrated in Mississippi, Arkansas, Alabama and Louisiana, while deer farms comprise significant farm revenues in Texas, Pennsylvania and Ohio. Over the 32 years of the Program, Land Grant institutions represented by the four Regional Coordinators have developed significant expertise in addressing the issues of regional producers. These same institutions have also dedicated facilities and personnel to meet the Good Laboratory Practices regulations (CFR 21.58).

Second, FDA/CVM approval of drugs requires performing clinical field trials in several geographical locations throughout the US. The rationale behind this requirement is that climate and other environmental conditions as well as possible differences in husbandry may affect the efficacy of any particular drug regimen. Coordinating these field trials spread throughout the US is most cost-effective through the multistate project approach. Our Regional Coordinators have worked closely together during the life of the Program to facilitate such work. Projects such as our ivermectin cattle tick study in Texas is currently run through both Iowa State University and the University of Florida. Such coordination uses the ruminant expertise of Iowa State veterinarians with the analytical capabilities of the University of Florida.

Thus, only a multistate project could possibly address the needs of such a widespread collection of industries and Federal regulations in a cost-effective manner.

What the likely impacts will be from successfully competing the work - Since its inception in 1983, the Minor Use Animal Drug Program has been responsible for generating 52 New Animal Drug Applications, an average of 1.6 per year. For the 5-year period of this review, NSRP-7 was responsible for three PMF/drug approvals – Progesterone Solid Matrix for sheep (NADA 141-302), Lincomix for the control of American foulbrood in honey bees (NADA 111-636) and Chloramine-T for the control of mortality in freshwater-reared salmonids due to bacterial gill diseases (NADA 141-423). Only NADA 141-302 received Program funding during the latest 5-year period, as research supporting NADA 111-636 was conducted at USDA/ARS and data for NADA 141-423 was developed during the previous 5-year period. Further, the Program has supplied supplemental data to the US Fish and Wildlife Services in support of 21 New Animal Drug Applications. Together these PMFs have supported FDACVM approval for 73 drug products for use in minor food species.

Additionally, from inception, regional coordinators have published 211 peer-reviewed articles relating to the use of therapeutic drugs in minor species. Such publications benefit stakeholders by providing veterinarians with the necessary information to allow the extra-label use of these drugs on minor species.

NRSP-7 was established and is still needed today because there are insufficient financial incentives for the veterinary pharmaceutical industry to invest in minor species. Additionally, there are potential liability issues from animal injury in minor species that may create unfavorable risk-reward relationships for pharmaceutical companies that add these species to established label claims. NRSP-7 develops the data to support the drug approval process and eventually the addition of the minor species to the label claim. In doing so, NRSP-7 helps to improve the international competitiveness of US agriculture. As the veterinary pharmaceutical industry continues to undergo worldwide consolidation, the resources devoted to minor species and minor uses continue to diminish. This makes the role of NRSP-7 increasingly critical for maintaining the efficient and competitive production of these minor species and minor uses in the US.

Through a productive NRSP-7, producers and veterinarians will continue to have the necessary information to prevent disease-related losses, to reduce pain and suffering in important species, and to avoid contamination of our foods with drug residues and drug resistant pathogens.

### Rationale: Priority Established by ESCOP/ESS

The seven ESCOP National Priorities include: (1) Develop new and more competitive crop products and new uses for diverse crops and novel plant species; (2) Develop new products and new uses for animals; (3) Reduce the risks of local and global climatic change on food, fiber, and fuel production; (4) Provide the information and knowledge needed to further improve environmental stewardship; (5) Improve the economic return to agricultural producers; (6) Strengthen our communities and families and (7) Ensure improved food safety and health through agricultural and food systems.

NRSP-7 research addresses three of the seven ESCOP Roadmap Challenges, including Challenge 2, Challenge 5, and Challenge 7. The primary contribution of NRSP-7 is to *Ensure improved food safety and health through agricultural and food systems (Challenge 7).* Concern over drug residues in our food supply has grown exponentially over the last 10 years. Food producers and veterinarians are under pressure to limit the use of antimicrobials in food animals and employ more prudent oversight over even therapeutic uses. Prior to the initiation of the Program in 1982, private sponsors had supported approvals for the use of minor use drugs as follows: none for rabbits, one for ducks and pheasants (none for other game birds), two for food fish, four for goats and 21 for sheep. A majority of these approvals represented outdated drugs with insufficient data on the post-treatment residue levels as well as efficacy. In 32 years, data supplied by the Program provided for the publication of 52 New Animal Drug Applications and modifications per year or 2.3 new label claims per year. Included in these Public Master Files were one for rabbits, nine for game birds, 16 for fish, lobster and shrimp, 15 for meat and dairy goats, eight for bison and reindeer, one for foxes and two for honey bees.

In addition to adding minor species to label claims, the Program works to ensure food safety through the publication of data on the pharmacokinetics, safety and effectiveness of modern drugs in minor species. Extra-label use of drugs by veterinarians, requires knowledge of the pharmacokinetics, tissue distribution, and sensitivity of the animal to support the decision to treat with an unapproved therapeutic. Regional coordinators in the Program have published 211 peer-reviewed articles supporting veterinarians in their decision to use drugs in an extra-label

use manner. This information is also supplied to the FARAD program for online access to veterinarians (<u>http://www.farad.org</u>).

Thus, the linkage to human health includes improving the quality and safety of food, addressing issues of zoonotic diseases that threaten both animal and human health, and assuring safe and efficacious animal health products that do not adversely affect human health.

Additionally, with respect to (*ESCOP Roadmap Challenge 5*), *Improve competitiveness and profitability/economic return to the producer in agriculture*, NRSP-7 serves as a critical support component of minor species production systems in the US. Economic survival of these minor species producers depends upon their ability to treat diseases with approved drugs and the knowledge that such treatment will not harm the species or incur illegal drug residues. Production units or farms for minor species typically operate on thin margins. Economic success or even survival depends on optimal health of the crop. Limiting disease with approved therapeutics allows producers to treat animals with confidence and assurance that the dose selected will perform as intended and will not result in illegal contamination.

The study of ivermectin medicated feed blocks for cattle fever ticks by NRSP-7 is another example in which the program supports improved competitiveness in agriculture. Treatment of cattle with ivermectin-treated molasses blocks within the 852-square mile quarantine zone that runs along the Mexico-Texas border was identified as a minor use in a major species. Thus, the experience of the Program with the analysis of ivermectin and the drug approval process has enabled NRSP-7 to establish pivotal efficacy and safety data and await only the cooperation of the manufacturers to put the newly documented products to use in the quarantine zone. Since most other treatment options have failed, without these new ivermectin/molasses blocks in use, the entire U.S. cattle industry is at risk if cattle tick fever returns to the United State (www.angusbeefbulletin.com/extra/2008/dec08/images/.../texas\_tick.pdf).

Work by NRSP-7 led to FDA/CVM approval for progesterone implants for estrus synchronization in sheep. This has enabled sheep farmers to begin producing lambs throughout the year. This in turn allows the producer to supply animals to the market at times other than peak "natural" breeding periods, improving US sheep farming competitiveness in the US and foreign markets.

NRSP-7 also assists to **Develop new products and new uses for animals** (Challenge 2). Without NRSP-7, the introduction of many new animal species would not be possible since they are generally high value specialty animals with few, if any, approved therapeutics. The work on the development of the ivermectin/molasses blocks described in the context of Challenge 5 is also consistent with Challenge 2 and is discussed in more detail above.

#### **Rationale: Relevance to Stakeholders**

Animal producers are the primary stakeholders in the NRSP-7 program, but pharmaceutical companies may be considered significant stakeholders as well. Other groups with interest in minor animal drug use include veterinarians and regulators. The active participation of animal producers and pharmaceutical companies is essential for the success of the program. However, to one degree or another, NRSP-7 involves all stakeholders. NRSP-7 producer stakeholders are represented by the following 58 organizations in 10 categories: American Association of Wildlife Veterinarians, American Association of Zoo Veterinarians, American Farm Bureau, American Feed Industry Association, American Pet Product Manufacturers Association, Inc., American Rabbit Breeders Association, American Sheep Industry Association, American Veterinary Medical Association, Animal Health Institute, Animal Drug Alliance, Arkansas Bait and Ornamental Fish Growers Association, Inc., Food Animal Concerns

Trust, International Association of Aquatic Animal Medicine, International Association of Fish and Wildlife Agencies, North American Deer Farmers Association, North American Gamebird Association, Inc., National Pork Producers Council, National Cattlemen's Beef Association, National Fisheries Institute, National Turkey Federation, Pacific Coast Shellfish Growers Association, and the National Aquaculture Association.

By category, stakeholders include:

- **Agencies** Fish and Wildlife Service, and Animal and Plant Health Inspection Service, US Geologic Survey, and FDA Center for Veterinary Medicine.
- Aquaculture: Marine Fish Dealers, American Tilapia Association, U.S. Trout Farmers Association, American Fisheries Society, World Aquaculture Society, Aquaculture Network Information Center, and Catfish Planet.
- **Bees**: American Beekeepers Federation, American Honey Producers Association, International Bee Research Association, Iowa State Entomology Index: Beekeeping, and Beekeeper's Home Pages Internet Resources.
- **Game Birds**: Mississippi State Game Bird Management, Pheasants Forever, Quail World, and North American Game Bird Association.
- **Caprine**: American Dairy Goat Association.
- **Institutions**: Auburn University (aquaculture and fisheries), Cornell University (aquaculture), Minnesota (avian), and Texas AMU (poultry science).
- Lagomorphs: American Rabbit Breeders Association.
- **Ovine:** American Sheep Industry Association
- **Ratites**: American Emu Association and The American Ostrich Association.
- **Reptiles**: The Gator Hole and Crocodilian Internet Resources.
- **Ungulates**: Alpaca Registry, National Bison Association, The White-Tailed Deer Farmer's Network, North American Deer Farmers Association, Deer Hunting Net, and North American Elk Breeders Association.

These stakeholders provide input to NRSP-7 as to their individual drug needs and support projects through the contribution of animals, facilities for drug testing, commercial drugs, data, and analytical methodology.

*Stakeholder needs* – Veterinary medicine has a key role in protecting the health and productivity of several billion farm animals worldwide, ensuring the quality of the food they yield, and in protecting the health of approximately one billion companion animals. Even though a large number of animals are treated with veterinary pharmaceuticals, the human healthcare market is about 35 times larger than the combined market for all non-human species, which had a global value in 2012 of \$21.1 billion. This figure can be divided among three main modalities: veterinary pharmaceuticals, biologicals and medicated feed additives (MFAs). 63% of the \$21.1 billion veterinary health market in 2011 is accounted for by pharmaceuticals (\$13.3 billion), 25% by biologicals (\$5.27 billion) and 12% by medicinal feed additives (\$2.5 billion). Food animal healthcare comprised \$12.45 billion of the total 2011 market; companion animal healthcare comprised \$12.45 billion of the total 2011 market; animal healthcare comprised \$8.64 billion (http://finance.yahoo.com/news/global-veterinary-health-products-market-193000008.html). Moreover, this animal market is dominated by a large number of products that generate small revenues, and so the balance between the amount of R&D investment required relative to the likely return on this investment is a particularly crucial issue in veterinary drug development.

Minor species represent an excellent example of this disparate market. Agricultural production of fish, gamebirds, sheep, goats, ratites bees and deer in the United States is critically important to numerous regional economies in the United States. This diverse aggregation of minor species represents approximately \$4.8 billion in state and local US farm revenues annually.

Additionally, processing and export of minor species food and fiber products represents an additional \$36.6 billion of revenue. Individually, however, these minor species represent drug markets too small to provide a sufficient return on the high cost of developing a new animal drug application.

Prior to NRSP-7, the FDA/CVM had approved the use of drugs for minor species as follows: none for rabbits, one for ducks and pheasants (none for other gamebirds), two for food fish, four for goats and twenty-one for sheep (most of which have been withdrawn). Minor and specialty use needs have continued to accumulate, leaving the producer of these species without the drugs necessary for disease prevention and control. The Minor Use Animal Drug Program has received 354 Animal Drug Requests submitted by animal producers, researcher investigators at federal, state, and university laboratories, veterinarians, and animal industry personnel for approval of a specific drug for the control of a certain disease in a particular animal industry. Of these requests, more than 40 have been identified as priority projects for NRSP-7.

*Measuring stakeholder use* – Animal produces who use unapproved drugs for the treatment of livestock face the liability of illegal drug residues as well as the risk of ineffective dosages. Before the Minor Use Animal Drug Program, these producers had little choice, but to use unapproved drugs when faced with outbreak situations. Without these drugs, animal suffering and mortality would greatly increase, as would the cost of producing animal-derived food product. The FDA/CVM is aware that veterinarians and livestock producers were using unapproved drugs without the safeguards that approved drugs carry. Because there is widespread use of unapproved drugs in minor species and the level of use is small, approval of drugs for the minor use needs does not generally result in a measurable increase in sales to the pharmaceutical company. Thus, it is not possible to achieve a measure of stakeholder use of NRSP-7 data and drug approvals through increases in drug sales. One major exception to this situation is in the area of aquaculture. Increases in sales and usage of NRSP-7 developed drug approvals can be monitored through medicated feed records and pharmaceutical company shipment records.

Two robust measures of stakeholder use include the conversion of published Public Master Files to New Animal Drug Applications and the addition of the minor use on the label of the pioneer product for the pharmaceutical stakeholder and the "in-kind" contributions of producers in supporting the research of NRSP-7. Of the 32 Public Master Files supported entirely by NRSP-7 research, 52 label claims have been added to products for minor uses. Additionally, a total of 73 label claims have been developed containing data generated by NRSP-7. Thus, pharmaceutical companies have been strongly supportive of the Program and taken advantage of the opportunity to support the prudent use of antibiotics even though such action increases their liabilities out of proportion to profits. The second metric of stakeholder support, in-kind contribution of resources and facilities, has accounted for an estimated \$982,800 over the last five years of the Program (see Appendix F).

#### **Objectives and Projected Outcomes**

The objectives of NRSP-7 are:

- 1. Identify the animal drug needs for minor species and minor uses in major species.
- 2. Generate and disseminate data for the safe, effective, and legal use of drugs intended for use in minor animal species.
- 3. Facilitate FDA/CVM approvals of drugs for minor species and minor uses.
- 4. Provide alternatives to antibiotic use in food animals through the identification and FDA/CVM approval of naturally occurring biotherapeutics and feed additives.

#### Objective 1 - Identify the animal drug needs for minor species and minor uses in major species

Critical and emerging needs are identified by the Minor Use Animal Drug Program Technical Committee based on information obtained from stakeholders represented by animal industry groups and producer organizations, scientific and professional groups, literature surveillance, and research originating within the program. To further refine specific project objectives, contacts are made with key, knowledgeable representatives from producer organizations, scientific and professional groups, government agencies, and pharmaceutical companies. Identification of producer group workshops and symposia proceedings are also used to gather information about priority needs and emerging issues.

The highest priority is given to research projects that address drugs or compounds that are required to prevent or treat disease, or for reproductive management. Drugs intended for the prevention or treatment of disease or for the modification of specific physiological functions in minor species, or to treat or prevent minor diseases in major food animal species are selected over drug requests for nonfood animals.

Specifically, a system has been devised to review, evaluate and recommend the feasibility of each animal drug clearance proposal submitted. When a proposal is accepted, the necessary data are obtained, compiled and submitted to FDA/CVM for establishment of a public master file, which will lead to the approval of the drug.

The process for selection of drugs for testing in NRSP-7 is represented schematically in Figure 1. Filing of an Animal Drug Request (ADR) form by any group or individual associated with specialty animal production initiates the process. Representatives of such groups include animal producers or their representative organizations, pharmaceutical manufacturers, university faculty and veterinarians. An example of the information requested for an ADR is provided in Appendix A. This ADR request form can be submitted online at <u>www.NRSP7.org</u> or through any of the four Regional Drug Coordinators, or the National Coordinator., Once received, the ADR is assigned a unique ADR number and included in the master ADR listing maintained at FDA/CVM, the National Coordinator's headquarters and at <u>www.NRSP7.org</u>.

During the spring annual meeting the NRSP-7 Technical Committee and representatives of the Stakeholder Advisory Committee (SAC) review the current projects and consider new ADR for funding. Each newly received ADR is then evaluated by the Technical Committee and SAC according to established criteria that include (1) availability of a pharmaceutical manufacturing sponsor, (2) major species approval, (3) microbial resistance concerns, (4) significance to the animal industry, (5) cost of developing the necessary data, and (6) food safety implications. ADR requests that meet these criteria are considered as potential projects.

Specific regional coordinators are assigned follow-up of all potential projects generally decided by regional expertise. Further concerns regarding the potential project are then addressed including: (1) the identification of researchers and research facilities, (2) development of FDA/CVM approved protocols with reasonable numbers of animals, and (3) scheduling. Monthly conference calls of regional coordinators, administrative advisors, FDA/CVM and USDANIFA liaisons provide continued follow-up of potential project progress.

Regional Coordinators determine: (1) what kind and how much work has been done on the compounds selected for study, (2) the approval requirements, (3) data collection capabilities available at the leader laboratory and at other laboratories in the region, (4) level of funding required, (5) whether a proper field research program is underway that will provide samples for analysis, and (6) initiate negotiations for such financial support as may be needed for performance of necessary work at other universities, federal agencies, or private concerns.

Where possible and appropriate Regional Coordinators will work closely with the National Coordinator in establishing proposed priority lists for projects, in negotiating for laboratory and field work that may be performed in their regions, and in delegating authority to them for conducting sub-projects funded in their regions. The Regional Coordinator may award "grants-in-aid" or contracts for field and laboratory work necessary to obtain the required data for registration petitions. Decisions to conduct specific projects are based on unanimous approval by the Technical Committee of the Animal Drug Program. Other experts will be consulted as deemed necessary to help with prioritizing projects.

Finally, in order to ensure that the program is responding to developing needs of minor species, research priorities are continually updated through review of producer group workshops and meetings. Weighted algorithms have been used to provide a general ranking or prioritization of requests. It has been found, however, that weights may change from year to year and the best process is continued, annual evaluation of stakeholder priorities.

# Objective 2 - Generate and disseminate data for the safe, effective, and legal use of drugs used in minor species or limited use in major species.

The following outlines the steps undertaken to initiate the generation and dissemination of data:

2.1 All projects are initiated by the submission of an Animal Drug Request (ADR) form to the National or a Regional Animal Drug Coordinator. The ADR's are forwarded to the FDA/CVM to request comments regarding the extent of the data package that would be required for drug/compound approval (e.g., target animal safety, effectiveness, residue depletion, and/or environmental assessment studies-see below). The appropriate pharmaceutical sponsor in cooperation with the efforts of NRSP-7 will seek to gain FDA/CVM approval of the drug formulation. Pending favorable initial review by both the sponsor and FDA/CVM, the ADR is ranked according to priority for the funding by the program. Upon receipt of the reviews from FDA/CVM and the pharmaceutical sponsor, a decision can be made to fund the project. The project objectives may be directed toward generating sufficient data to seek FDA/CVM approval of the drug (Objective #3), or when that is not practicable, toward generating data sufficient to support safe, effective and legal use under the AMDUCA legislation through publication in peer reviewed journals. Research not conducted in the laboratories of the Regional Animal Drug Coordinators is conducted under subcontracts, managed by the Coordinators, to scientists in qualified laboratories at other institutions.

(1) A product development meeting is then held for the participating Regional Animal Drug Coordinator(s) and FDA/CVM to identify specific data requirements needed to seek approvals. Investigators under subcontract with approved projects must prepare detailed research protocols that fulfill these requirements under the guidance of the Regional Animal Drug Coordinators, pharmaceutical sponsor and FDA/CVM.

(2) Following research protocol review and refinement with FDA/CVM and the pharmaceutical sponsor, the research may begin. Studies typically required in this phase include effectiveness (is the drug effective and at what dose), target animal safety (toxicity), human food safety (drug residues in edible products), and environmental assessment (as required).

(3) Upon completion of the required studies, formal reports of the required studies are submitted to FDA/CVM for evaluation. Upon completion of an approved study, these data are published at the FDA/CVM site in either phase review form or completed data package. Once publicly available, the manufacturer may use this information by reference to support a new label claim (NADA or ANADA). Another form of dissemination of data occurs when the researcher publishes the results of the research in a peer-reviewed article.

2.5 In 32 years, data supplied by the Program provided for the publication of 32 Public Master Files supporting 52 New Animal Drug Applications and modification of 73 label claims to include minor species, an overall average of 1.6 Public Master Files per year or 2.3 new label claims per year. Included in these New Animal Drug Applications were one for rabbits, nine for game birds, 16 for fish, lobster and shrimp, 15 for meat and dairy goats, eight for bison and reindeer, one for foxes and two for honey bees.

2.6 In addition to adding minor species to label claims, the Program provides information through the publication of data on the pharmacokinetics, safety and effectiveness of modern drugs in minor species. Extra-label use of drugs by veterinarians, requires knowledge of the pharmacokinetics, tissue distribution, and sensitivity of the animal to support the decision to treat with an unapproved therapeutic. Regional coordinators in the Program have published 211 peer-reviewed articles supporting veterinarians in their decision to use drugs in an extra-label use manner. This information is also supplied to the FARAD program for online access to veterinarians (<u>http://www.farad.org</u>).

#### Objective 3 - Facilitate FDA/CVM approvals of drugs for minor species and minor uses.

Upon completion of each required study, reports of results and all raw data are submitted to the Regional Animal Drug Coordinator for review prior to submission to FDA/CVM. Following acceptance of the data by FDA/CVM, a Public Master File is established that is placed in the public domain (e.g., published on the CVM website). The pharmaceutical sponsor may refer to the Public Master File in support of a New Animal Drug Application (NADA), and ultimately the labeling of the drug for the use in the minor species, or for the minor use in a major species.

NRSP-7 functions through the coordination of efforts among animal producers, pharmaceutical manufacturers, FDA/CVM, USDA/Cooperative State Research, Education, and Extension Service, universities, state agricultural experiment stations and veterinary medical colleges though out the country. The steps involved in this coordination of efforts are presented schematically in Figure 1.

# Objective 4 - Provide alternatives to antibiotic use in food animals through the identification and FDA/CVM approval of naturally occurring biotherapeutics and feed additives.

Worldwide, government agencies and the public have become concerned about the continued use of antibiotics in food producing animals. Scientists around the world have provided strong evidence that antibiotic use in food-producing animals can have a negative impact on public health through the following sequence of events: (1) Use of antibiotics in food-producing animals allows antibiotic-resistant bacteria to thrive while susceptible bacteria are suppressed or die; (2) Resistant bacteria can be transmitted from food-producing animals to humans through the food supply; (3) Resistant bacteria can cause infections in humans; and (4) Infections caused by resistant bacteria can result in adverse human health consequences. [http://www.cdc.gov/narms/animals.html] Because of the link between antibiotic use in food-producing animals and the occurrence of antibiotic-resistant infections in humans, the FDA and CDC encourage continuing efforts to minimize inappropriate use of antibiotics in humans and animals. [http://www.fda.gov/AnimalVeterinary/%20SafetyHealth/AntimicrobialResistance/JudiciousUseofAntimicrobials/default .htm]

The FDA has taken the position to promote the judicious use of antibiotics that are important in treating humans. This strategy recommends that such antibiotics should be used in food-producing animals only under veterinary oversight and only to address animal health needs, not to promote growth. The American Veterinary Medical Association has, in concert, developed a position statement on the judicious therapeutic use of antimicrobials, essentially limiting any use to under the direct supervision of a veterinarian.[https://www.avma.org/KB/Policies/Pages/Judicious-Therapeutic-Use-of-Antimicrobials.aspx']

<u>5. Expected Outcomes</u> - Data generated through this project will lead to improved animal health and welfare (reduction of pain and suffering) as new applications of drugs for minor species are made available. Many drugs are currently used without appropriate labeling or without an environmental impact assessment of the drug (e.g. some aquaculture drugs and most drugs for all *Cervidae*). In addition, the availability of scientific data supporting proper dose and duration of treatment will reduce the likelihood of antimicrobial resistance developing with implications for human health. The availability of additional approved drugs will also result in reduced economic losses for minor species producers.

Methods for communicating results will include public master files, approved product labeling, peer-reviewed publications in journals, abstracts presented at meetings, publications in producer/trade journals, presentations at workshops, symposia, producer group meetings, and continuing education programs. The beneficiaries of this project include the animals (health and welfare), animal producers (economic), and consumers (safety and economics) of the animal products through the promotion of human health, food safety and environmental protection.

# 5.1 Expected Outcome of Objective 1 - Identify the critical needs of the various producers of minor livestock species

To date 354 drug requests have been submitted by stakeholders to the Minor Use Animal Drug Program for the development of data in support of the submission of a New Animal Drug Application (NADA). Through a prioritization process that has included (i) constraints imposed by concerns of antimicrobial resistance, (ii) limitations of availability of certain expensive or rare animal species, (iii) appropriate efficacy models, and (iv) high risk/benefit liabilities and lack of economic incentives for pharmaceutical manufacturers, the number of highest priority projects has been estimated at 40. Budget cutbacks have reduced our current active projects to nine, with active research limited to 3 to 5 per year.

The Southern Region has taken responsibility for the NRSP-7 Home-Page [www.nrsp-7.org]. This resulted in reworking the public sector and, the IP limited access site ["Ringer Site"] that continues to allow members of the committee access to archival data, relevant media material, and information concerning on-going projects. The latter includes an ASP interactive database ["MUMS Rx"] that is available for public access.

Over the last six years, drug coordinators and the FDA/CVM liaison group have attended regular monthly teleconferences. These have been coordinated by the National Coordinator and have proved very successful in facilitating communication and coordination among the parties participating. These teleconferences usually take place at noon EST on the first Tuesday of each month. Additionally, twice each year, the Program conducts an annual teleconference meeting as travel costs have prohibited meetings in Washington, D.C.

# 5.2 Expected Outcomes of Objectives 2 and 3 - Generate and disseminate data for the safe, effective, and legal use of drugs used primarily in therapy or reproductive management of minor animal species. Facilitate FDA/CVM approvals of drugs for minor species and minor uses.

NRSP-7 has published 211 articles in peer-reviewed journals, averaging 6.6 per year over the term of the program. For the last five years, however, publications have increased to nearly 10 per year. Thus, although FDA/CVM drug approvals have waned due to increasing costs, the Program has increased its efforts to supply critical data needs to minor species producers. The data generated by the Program is also shared with the Food Animal Residue Avoidance Database (FARAD program to further increase visibility. For a detailed description of NRSP-7 dissemination of data and information, see Outreach and Communication.

Since its inception in 1983, the Minor Use Animal Drug Program has been responsible for generating 32 Public Master Files supporting 52 New Animal Drug Applications, an average of 1.6 New Animal Drug Applications per year. Included in these approvals were one for rabbits, nine for game birds, 16 for fish, lobster and shrimp, 15 for meat and dairy goats, eight for bison and reindeer, one for foxes and two for honey bees (Table 2).

For the 5-year period of this review, NSRP-7 was responsible for three Public Master Files – Progesterone Solid Matrix for sheep (NADA 141-302), Lincomix for the control of American foulbrood in honey bees (NADA 111-636) and Chloramine-T for the control of mortality in freshwater-reared salmonids due to bacterial gill diseases (NADA 141-423). Further, the Program has supplied supplemental data to the US Fish and Wildlife Services in support of 21 New Animal Drug Applications. Together all these Public Master Files have supported FDACVM approval for 73 new drug products for use in minor food and fiber species. Currently there are nine active research projects involving five animal species and seven different drugs (Table 2). Ruminant species remain the predominant group with a majority of Public Master Files (53%) as well.

# 5.3 Expected Outcomes of Objective 4 - Provide alternatives to antibiotic use in food animals through the identification and FDA/CVM approval of naturally occurring biotherapeutics and feed additives.

Modeling after the IR-4 Program's work with HopGuard for varroa mites in honey bees, NRSP-7 has initiated discussion with BetaTech for inclusion of their hops beta acids into the Program. NRSP-7 would assess the potential of these beta acids to serve as a substitute for antibiotics as growth promoters in animal feeds. BetaTech currently holds four patents on a number of hops acids as growth promoters and bioterapeutics for a range of bacterial infections in ruminant and avian species. Discussions with FDA/CVM have been positive and the Program looks forward to beginning to work with game bird stakeholders in assessing the potential of these natural compounds. As feed additives and growth promoters, the hops beta acids will fall under different FDA/CVM regulations than therapeutics, but the Agency has already assured NRSP-7 it will work to accommodate these changes.

Objective 4 would also provide a more direct visibility of the Program's efforts to support prudent use of antibiotics in food producing animals. This additional objective will draw more support from Congressional members who, in the past, have been reluctant to unconditionally support the Program for fear this support would be interpreted as increasing antibiotic use in animals. As previously detailed, the public is keenly aware of the issue of antibiotic resistance and efforts to reduce the use of antibiotics in food producing animals are strongly supported.

#### Management, Budget, and Business Plan

<u>1. Organizational structure</u> - NRSP-7 is composed of a Technical Committee and four Administrative Advisors representing state experiment station directors. These Administrative Advisors provide liaison among the directors of the state experiment stations, USDA/NIFA, FDA/CVM, various animal organizations, and others coordinating the efforts of this program.

The organizational structure of the Minor Use Animal Drug program follows:

## Administrative Advisory Committee

The Administrative Advisory Committee is composed of one appointee by Experiment Station Directors from each of the four regions (North Central, Northeast, Southern, and Western). The chair of the committee is selected internally. The role of the Administrative Advisory Committee is to provide liaison among the directors of the agricultural experiment stations in the four regions, colleges of veterinary medicine, the USDA/NIFA, the FDA/CVM, various animal organizations, and with those coordinating the efforts of this program. This committee establishes and sets policy consistent with the mission of the project. This committee also advises on budget and administrative matters relating to the program.

#### Technical Committee

The Technical Committee is composed of the following representatives:

- National Animal Drug Coordinator (Chair)
- Regional Animal Drug Coordinators representing each of the four regions (North Central, Northeast, Southern, and Western)
- Administrative Advisory Committee Chair (non-voting)
- USDA/NIFA Representative (non-voting)
- FDA/CVM liaison to NRSP-7 (non-voting)

In addition to the above committee, the FDA/CVM has a group of Minor Use Animal Drug reviewers that meets with the Technical Committee generally once a year at the semi-annual meetings of the Technical Committee. This FDA/CVM group consists of representatives from the Division of Therapeutic Drugs for Food Animals, the Division of Human Food Safety, the Division of Production Drugs, and the Environmental Sciences Staff. The National Animal Drug Coordinator is salaried on a part-time basis and maintains an office. The Regional Animal Drug Coordinators are not compensated by salary except for secretarial or technical services.

Cooperating Agencies and Principal Leaders:

US Department of Agriculture/CRESS	
Dr. Gary Sherman	USDA/CRESS Representative
US Food and Drug Administration/Cente	r for Veterinary Medicine
Dr. Meg R. Oeller	FDA/CVM Liaison
Dr. Amy Omer	FDA/CVM Liaison
Dr. Dorothy Bailey	FDA/CVM Liaison
Administrative Advisors	
Dr. George Smith	Michigan AES
Dr. Margaret E. Smith (Chair)	New York AES
Dr. Frances D. Galey	Wyoming AES
Dr. Philip H. Elzer	Louisiana AES
National Coordinator	
Dr. John G. Babish	New York AES
Regional Coordinators	
Dr. Lisa Tell	California AES
Dr. Rodman G. Getchell	New York AES
Dr. Thomas Vickroy	Florida AES
Dr. Ronald W. Griffith	Iowa AES

#### 2. Funding activities

In the past, Research for the Minor Use Animal Drug Program was funded through a USDA special research grant administered by NIFA in cooperation with the NRSP-7 Technical Committee. Currently, however, NRSP-7 has been dependent on "off-the-top" Regional Research funds allocated to the Minor Use Program. Table 3 below summarizes the five-year funding of NRSP-7 the Minor Use Animal Drug Program (MUADP), for the years 2009 – 2013 inclusive. This period represents the previous NRSP approval term of the MUADP. Total funding from all sources including Cash and In-Kind was \$9.7 million or approximately \$2 million per year.

NRSP-7 is a cooperative program involving experiment stations, veterinary colleges, regulatory agencies, animal producers and pharmaceutical companies. FDA/CVM cash and in-kind support was \$1.8 million or 1.9-times Hatch funding received during this period. Support by

Stakeholders (producers and pharmaceutical companies) during this period was \$5.7 million or 6-times all Hatch funding. All outside support totaled \$9.7 million or 9.2-times the Hatch funding received by the MUADP over five years.

Overall, during the 2009-2013 period the NRSP-7 was largely supported by stakeholders with both cash and in-kind funding, while Hatch funding accounted for approximately 10% of the operational budget. This 10%, however, was essential to coordinate the necessary resources to achieve the level of productivity exhibited during this five-year period for our stakeholders.

Drug approvals are generally species and disease specific and additional label claims come with considerable added expense. Pharmaceutical company estimates place the cost of simply adding a label claim to a previously approved drug at \$10 to \$25 million. In the 2009-2013 period, NRSP-7 research made possible the addition of five new drug uses in minor species and successfully completed the human food safety study for fenbendazole in pheasants. For the 5year period of this review, NSRP-7 was responsible for three Public Master Files/drug approvals These wre Progesterone Solid Matrix for sheep (NADA 141-302), Lincomix for the control of American foulbrood in honey bees (NADA 111-636) and Chloramine-T for the control of mortality in freshwater-reared salmonids due to bacterial gill diseases (NADA 141-423). Only NADA 141-302 received Program funding during the latest 5-year period, as research supporting NADA 111-636 was conducted at USDA/ARS and data for NADA 141-423 was developed during the previous 5-year period. Further, the Program has supplied supplemental data to the US Fish and Wildlife Services in support of 21 New Animal Drug Applications. Together all these Public Master Files have supported FDACVM approval for 73 drug products for use in minor food and fiber species. Over the last five years, NRSP-7 cost for adding a minor species claim to a drug label was approximately 2- to 5-times more efficient than industry.

Perhaps the most significant of this "in-kind" support, however, comes through the cooperation of the pharmaceutical companies that provide access to their proprietary data package prepared for the drug approval in a major species, estimated at approximately \$100 million (http://www.ahi.org/about-animal-medicines/industry-statistics/). In addition, the pharmaceutical sponsors complete the approval package by adding the new use of the drug to their current label, and often contribute to the program in the form of providing the investigational drug for research, as well as direct financial aid. Without the generous support of the pharmaceutical manufactures, this program would not be possible.

The Regional Animal Coordinators are not compensated by salary for time contributed to the Minor Use Animal Drug Program. In two cases, secretarial and/or technical support services are budgeted from the Program. The funding of \$20,000 for the National Drug Coordinator's part-time salary (30%) and the maintenance of an office has been donated to program research by the National Coordinator over the last two years.

Overall, the mean total expenditure per completed research for a drug approval or publication of a Public Master File was \$668,089. Average federal expenditures per completed research for publication of a Public Master File was \$409,907. Moreover, with 73 additional label claims, the total federal cost per label claim generated from NRSP-7 research has been \$185,300. The process of generating the safety and efficacy data necessary for FDA approval of a drug is costly and time-consuming. At present, the estimated cost to a pharmaceutical company for research necessary to obtain FDA approval for a new drug exceeds \$80 to \$100 million, and requires 8 to 10 years of concentrated research effort. The addition of a new label claim is also costly, ranging from \$10 to \$25 million.

Even with the estimated increased cost per drug approval in recent years, the NRSP-7 program continues to demonstrate remarkable efficiency and cost effectiveness. Compared to an average investment of the pharmaceutical industry of \$10 to \$25 million for adding a label claim

to an existing veterinary drug, information generated for additional label claims by the NRPS-7 program costs only approximately 15 to 35% of pharmaceutical industry costs (http://www.ahi.org/about-animal-medicines/industry-statistics/).

#### 3. Research

Research projects are initiated by requests, usually from researchers or animal producers, to the program's regional coordinators to address a particular minor use animal drug need. These requests, known as ADRs (Animal Drug Requests), are prioritized according (i) to financial and regulatory feasibility, (ii) to importance to the animal industry, and the pharmaceutical manufacturer's commitment to the minor use animal drug approval. Once a request is accepted as a research project, study protocols are developed and reviewed by FDA/CVM. All research projects are conducted in accordance with FDA's Good Laboratory Practices regulations. This process is outlined schematically in Figure 1.

**Research Planned for Upcoming Year** - To date 354 drug requests have been submitted to the Minor Use Animal Drug Program for the development of data in support of the submission of a New Animal Drug Approval (see NRSP-7 website for a complete listing of drug requests). Through a prioritization process that has included (1) constraints imposed by concerns of antimicrobial resistance, (2) limitations of availability of certain expensive or rare animal species, (3) appropriate efficacy models, and (4) high risk/benefit liabilities and lack of economic incentive for certain pharmaceutical manufacturers, the number of highest priority projects has been estimated at approximately 40. Of these, the Program has been actively working on nine projects.

Over the last five years the total Federal plus non-Federal cost for NRSP-7 to provide the data necessary to support a single label claim has risen to approximately \$3.5 million. This increase is due to (1) more sophisticated analytical procedures for residue analysis, (2) the need to conduct all studies under Good Laboratory Practices and auditing of projects, and (3) more expensive environmental assessments. Federal costs per Public Master File are estimated at roughly half this amount or \$1.75 million. With NRSP-7 total level of funding of approximately \$300,000 per year and cost per drug approval of \$1.75 million, the expected time for achieving a drug approval is 5.6 years. Thus, it is anticipated that NRSP-7 will achieve one approval over the next five years.

This level of progress falls critically below the needs and expectations of our stakeholders and it is the objective of the Program to use the next year to evaluate the continued viability of the program in the face of continuing escalating costs and dwindling funding.

<u>3.1 Research planned</u> - Over the next five years, the Program will work to organize on several fronts to establish the potential for increased funding. First, the Program will work to obtain more grants from the FDA/CVM Minor Use Minor Species (MUMS) grant program. Since the inception of the MUMS program at the FDA/CVM, NRSP-7 has taken advantage of this funding for several projects, but can increase this number in the upcoming year. This year I-010536 Strontium Chloride for otolith marking study has applied for FDA MUMS funding to supplement the Program's Hatch Funding.

Second, NRSP-7 will critically review the current nine active projects with the goal of completing two or three of the most visible. Projects likely to be completed in the coming year include I-006013 Erythromycin in Salmonids, I-011389 CIDR implants for goats, I-010062 Fenbendazole in pheasants, and I-012056 Ivermectin block for cattle tick fever.

Additionally, the Program will solicit support from companies interested in developing markets for natural alternatives to antibiotic growth promoters. For example, BetaTech of Washington, DC, has developed data on the growth promoting and antibacterial properties of hops beta acids. In US patent 7,090,873 the growth promoting properties of these alpha acids on cattle were

demonstrated. The Program has had discussions with BetaTech for interest in developing these applications for both major and minor species and the Company has been favorable to working with NRSP-7.

#### 4. FDA/CVM approval

A successful research project is submitted to FDA's Center for Veterinary Medicine for review and inclusion in a Public Master File. The availability of the data for use on a label claim is announced through publication of the Public Master File in the *Federal Register*. A pharmaceutical sponsor may then reference, at no cost, the data in the Public Master File to support a new animal drug application for the minor use. The final step in the process is FDA/CVM approval of this application for the pharmaceutical sponsor, so that the product may be labeled and sold.

#### 5. Assessment of outcomes

*Productivity* - In the 32 years, data supplied by the Program provided for the publication of 32 Public Master Files, 52 New Animal Drug Approvals and modification of 73 label claims to include minor species, an overall average of 1.6 New Animal Drug Approvals per year or 2.3 new label claims per year. Included in these New Animal Drug Approvals were one for rabbits, nine for game birds, 16 for fish, lobster and shrimp, 15 for meat and dairy goats, eight for bison and reindeer, one for foxes and two for honey bees.

Additionally, NRSP-7 has published 211 articles in peer-reviewed journals, averaging 6.5 per year over the term of the program. For the last five years, however, publications have nearly doubled to 11.4 per year. Thus, although FDA/CVM drug approvals have waned due to increasing costs, the Program has increased its efforts to supply critical data needs to minor species producers. The data generated by the Program are also shared with the Food Animal Residue Avoidance Database (FARAD( program to further increase visibility and assist veterinarians.

Currently there are nine active research projects involving five animal species and seven different drugs (Table 2). Ruminant species remain the predominant group with a majority of Public Master Files (53%).

NRSP-7 has also provided information on therapeutics in minor species use through peerreviewed publications, workshops and presentations to stakeholders and at professional meetings. Use of the Internet to optimize communications with stakeholders and program participants continues to improve in this rapidly changing medium. Moreover, NRSP-7 is the only initiative that generates information on the safe and effective use of therapeutics in minor species. Through NRSP-7, producers and veterinarians have the necessary information to reduce pain and suffering in commercially important minor species.

*Completion of original objectives* – A primary objective of NRSP-7 was to identify the animal drug needs for minor species and minor uses in major species. The Minor Use Animal Drug Program has received over 354 Animal Drug Requests submitted by researcher investigators at federal, state, and university laboratories, veterinarians, and animal industry personnel for approval of a specific drug for the control of a certain disease in an animal industry. Of these drug requests, the NRSP-7 Technical Committee has identified 40 of high priority. While in one sense NRSP-7 has completed one of our original objectives, withdrawal of available products, antimicrobial resistance, disease prevalence, husbandry practices, and the changing business relationships in the veterinary pharmaceutical industry preclude considering our current list of projects and potential projects as final.

The Program has generated and disseminated data for safe and effective minor species uses and minor uses in major species through the publication of 211 articles in peer-reviewed journals, averaging 6.6 per year over the term of the program. For the last five years, however, publications have increased to nearly 10 per year. Another form of dissemination of NRSP-7 data is the publication of drug pharmacokinetic and residue depletion studies through FARAD (Food Animal Residue Avoidance Database). FARAD is a computer-based decision support system designed to provide livestock producers, extension specialists, and veterinarians with practical information on how to avoid drug, pesticide and environmental contaminant residue problems.

With respect to facilitating FDA/CVM approvals for the above identified needs, In 32 years, the Program has provided data for the publication of 52 New Animal Drug Approvals and modification of 73 label claims to include minor species, an overall average of 1.0 Public Master File per year or 2.3 new label claims per year. For the 5-year period of this review, NSRP-7 was responsible for three Public Master Files. They included Progesterone Solid Matrix for sheep (NADA 141-302), Lincomix for the control of American foulbrood in honey bees (NADA 111-636) and Chloramine-T for the control of mortality in freshwater-reared salmonids due to bacterial gill diseases (NADA 141-423). Only NADA 141-302, however, received Program funding during the latest 5-year period, as research supporting NADA 111-636 was conducted at USDA/ARS and data for NADA 141-423 was developed during a previous 5-year period.

### Integration

Program facilitation and coordination exists among animal producers, pharmaceutical manufacturers, FDA/CVM, USDA/NIFA, other government agencies, state agricultural experiment stations, and schools of veterinary medicine. Animal producers have provided the majority of drug requests and they frequently supply animals and facilities for target animal safety and residue depletion studies. Pharmaceutical companies participate through the sharing of analytical methodology and providing commercial drug product for testing (See Funding). The major contribution of the pharmaceutical manufacturer is, however, is the cost borne for the approval of drugs for a major species, approximately \$100 million, and the cost of adding a label claim at \$10 to \$25 million (http://www.ahi.org/about-animal-medicines/industry-statistics/).

Since the beginning of the Minor Use Animal Drug Program, The FDA/CVM has supplied a fulltime liaison to coordinate the drug approval process. In the last four years, they have added part of the time of two additional staff positions to support the Program. The four regional coordinators are associated with colleges of veterinary medicine or experiment stations. These coordinators have full responsibility for supervising the development of all data entering the Public Master Files. Regional Coordinators also present and publish results of their studies. USDA/NIFA provides two full time liaison personnel who, along with the Administrative Advisors and Technical Committee, oversee the prioritization of drug requests as well as project planning and implementation. Persons serving as Administrative Advisors are provided by the agricultural experiment stations from the four regions of the United States. The fundamental need for NRSP-7 to operate as a functionally integrated program has existed since its inception and NRSP-7 has spent 32 years cultivating the relationships necessary for optimal efficiency.

## **Outreach, Communications and Assessment**

*Public Master Files and New Animal Drug Allowances (NADAs)* - The goal of the outreach and communication plan of NRSP-7 is to provide stakeholder access to information regarding program goals, accomplishments and impacts through a variety of channels. One form of outreach consists of the publication of the efficacy, target animal safety and drug residue depletion data generated as a Public Master File in the *Federal Register* and as a New Animal Drug Allowance (NADA) or Abbreviated New Animal Drug Allowance (ANADA). Publication in the *Federal Register* places the required studies in the public domain and a New Animal Drug Allowance provides the producer stakeholder the availability of the drug for the claimed minor

use. Table 2 lists the 33 Public Master Files and 52 New Animal Drug Allowances developed from data generated by NRSP-7.

Approvals and projects that have changed the outlook of two industries include the approvals of lincomycin hydrochloride water-soluble powder and tylosin tartrate powder for control of American Foulbrood in honey bees. These approvals represent a significant therapeutic addition to an industry working to reverse the declining honey bee population. Due to the small quantities of active ingredient required. the use of these products will not substantially increase sales of these drugs. The potential effect on the industry, however, is important. Secondly, the ivermectin/molasses block formulation study currently being conducted in Texas for the control of the cattle fever tick, has the potential of averting a major threat to the U.S. cattle population.

Several efficacy studies have supported the use of this novel formulation in cattle tick control. NRSP-7 labs in the Southern Region have worked with producers to assure a uniform distribution of ivermectin in the molasses blocks, insuring that cattle will receive uniform dosing.

*Presentations and publications* - Presentations, abstracts, publications and doctoral dissertations represent yet another form of communication to the stakeholders. Over the last 32 years, NRSP-7 has produced 211 peer-reviewed publications. Notably, while drug approvals have become more costly and time consuming to obtain over the last five years, NRSP-7 has nearly doubled its publication rate from 6.4 per year to 11.4 per year for the last five years. A listing of these publications follows this report.

*Website* - The Technical Committee has worked to develop the NRSP-7 website (www.NRSP7.org) as a communication tool for dissemination of information generated by the program. The site provides for the submission of Animal Drug Requests (ADR's), operational information and monitoring of project progress by Technical Committee members, access to the MUMS (Minor Use Minor Species) program and links to a variety of stakeholders' websites. The use of the Internet to optimize communications with stakeholders and program participants continues to improve in this rapidly changing medium. Since inception in 1999, the NRSP-7 website has been visited 13,251 times for an average of 2.4 hits per day. NRSP-7 believes that this represents a significant degree of interaction with stakeholders as well as the public at large.

Sharing NRSP-7 information with FARAD – Another form of dissemination of NRSP-7 data is the publication of drug pharmacokinetic and residue depletion studies through FARAD (Food Animal Residue Avoidance Database). FARAD is a computer-based decision support system designed to provide livestock producers, extension specialists, and veterinarians with practical information on how to avoid drug, pesticide and environmental contaminant residue problems.

The FARAD website (www.FARAD.org) provides:

- Current label information including withdrawal times of all drugs approved for use in food-producing animals in the United States.
- Official tolerances for drug and pesticides in tissues, eggs and milk.
- Database with approximately 43,000 scientific articles and entries with data on residues, pharmacokinetics and the fate of chemicals in food animals.

By supplying FARAD with information developed on minor use animal drug residue depletion and pharmacokinetics, NRSP-7 affords the stakeholder yet another conduit for obtaining critical information to avoid illegal and potentially hazardous drug residues in food animals.

*Steps to improve communications* – Several changes have been incorporated in an effort to enhance communication both within the program and with stakeholders. First, monthly teleconferences are held by the Technical Committee to discuss potential projects, interactions

with stakeholders and progress in studies. Second, stakeholders have been invited to be active non-voting participants in the annual spring teleconferences. The nature of the participation is *ad hoc* and representatives from different stakeholder groups are invited on a rotating basis, without representation from a single or specific group "assigned" to the committee.

Informatics have been better utilized to increase/improve communication with NRSP-7 participants and stakeholders. Improvements to web usage include posting pdf versions of publications and or dissertations that have been supported through NRSP7 funds as well as links to other appropriate pages (partners, producer and/or pharmaceutical company websites). Existing brochures and any newly developed media information packages are posted.

#### **Projected Participation**

NRSP-7 functions through the coordination of efforts among animal producers, pharmaceutical manufacturers, Food and Drug Administration/Center for Veterinary Medicine, United States Department of Agriculture/Cooperative State Research, Education, and Extension Service, universities, State Agricultural Experiment Stations and veterinary medical colleges throughout the country. Working relationships between the Program and both the FDA/CVM and NIFA have been and should continue to be excellent. Also, USDA/ARS has been participating with NRSP-7 in the cattle fever tick studies in Texas. Participation has also been forthcoming from game bird growers that have donated birds for safety and efficacy studies. Pharmaceutical companies have also provided analysis of feeds and tissues samples in selected studies. When the pharmaceutical companies could not provide analysis, they have provided expertise in the development of analytical methods for the tissue residue studies in minor species.

Further participation is projected from companies manufacturing natural products to be used as alternatives to antibiotics as growth promoters in animal feeds. An example of this is BetaTech of Washington, D.C. BetaTech already has its hops beta-acids in the IR-4 biopestices program under evaluation for control of the varro mite in honey bee hives. Initial discussions with BetaTech and FDA/CVM on supporting a project in NRSP-7 have been positive. FDA/CVM has indicated they will support NRSP-7 and BetaTech efforts to approve their hops beta acids as a growth promoting feed additive as part of its program to oversee the reduction of antibiotic use as growth promoters.

		US FARM GATE	US ECONOMIC
	LEADING	VALUE	IMPACT
INDUSTRY	STATES	[\$M]	[\$M]
Game Bird	TX, NC, PA, KS, WI, NY, IL, SD, FL,	\$897	\$5,401
	MN, IA, GA, MS, IN & AL.		
Rabbits	CA, GA, OH, PA, & TX	\$21.6	\$898
Honey Bees	ND, CA, SD, FL, MT, MN, TX, & WI.	\$166	\$17,284
Cervid	TX, PA, OH, FL, LA, IA, & KS	\$966 (farming)	\$3,241
		\$817 (hunting)	
Meat Goats	TX, TN, CA, GA, OK, NC, KY, MO, FL,	\$187	\$1,123
	& AL	\$205 (breeding)	
Dairy Goats	TX, OH, NY, PA, WI, WA, IN, CA, MD,	\$63.0	\$474
	MN, MI, FL, & KS.	\$16.0 (export)	
Sheep	TX, CA, WY & CO	\$810	\$4,861
Catfish/Aquaculture	Catfish	Catfish \$518	\$3,111
	MS, AK, AL, & LA	Trout \$94.6	\$172
	Trout		
	WA, WI, PA, ID, NC, OR, NY, CA, & CO		
	Total =	\$4,761	\$36,564

 TABLE 1.

 ECONOMIC IMPACT OF MINOR ANIMAL SPECIES BY STATE AS OF 2013

6	Ac:	TIVITY
INDUSTRY	APPROVALS (PMF#)	ACTIVE PROJECTS (INAD#)
Game Bird	Chukar partridges	Pheasants
	Sulfadimethoxine/Ormetoprim	Lasalocid (I-009096)
	(005-157)	Fenbendazole (I-010062)
	Lasalocid (005-429)	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
	Pheasants	
	Amprolium (003-887)	
	Thiabendazole (003-857)	
	Quail	
	Salinomycin (005-020) 2 NADA	
	Bacitracin (005-178)	
Rabbits	Monensin (005-014) 2 NADA Lasalocid (005-042)	
Foxes	Ivermectin (005-307)	-
Honey Bees	Tylosin (005-783) 3 NADA	
Holley Bees	Lincomycin (005-988)	
Cervid	Bison	
connu	Ivermectin (005-059) 4 ANADA	
	Reindeer	
	Ivermectin (003-895) 4 NADA	
Beef Cattle		Ivermectin (I-012056)
Meat Goats	Ivermectin (003-883)	CIDR (progesterone) (I-011389)
	Levamisole HCI (005-117)	Tulathromycin (I-011512)
	Albendazole (005-582)	
	Ceftiofur sodium (005-671)	
	Fenbendazole (005-118)	
	Monensin (005-055)	
	Decoquinate (005-012)	
	Morantel tartrate (005-366)	
Dairy Goats	Ivermectin (003-883)	CIDR (progesterone) (I-011389)
	Levamisole HCI (005-117)	Tulathromycin (I-011512)
	Ceftiofur sodium (005-671) Fenbendazole (005-118)	
	Monensin (005-055)	
	Decoguinate 005-012)	
	Morantel tartrate (005-366)	
Sheep	Bighorn Sheep	Sheep
50	Fenbendazole (005-071)	Florfenicol (I-011836)
	Sheep	Tulathromycin (I-011513)
	Decoquinate (005-258)	The Construction of the Construction of the State Construction of the
	Ceftiofur (005-544)	
	Tilmicosin phosphate (005-673)	
	CIDR (progesterone) (141-302)	
Catfish/Aquaculture†	Catfish	Fish
	Sulfadimethoxine/Ormetoprim	Erythromycin (I-006013)
	(005-056) Finfish	Strontium chloride (I-010536)
	Formalin (005-228) 9 NADA	
	Oxytetracycline (005-667) 4 NADA	
	Hydrogen peroxide (005-639)	
	Florfenicol (005-932)	
	Lobster	
	Oxytetracycline (005-028)	
	Shrimp	
	Formalin (005-228)	

## Table 2 Animal Drug Approvals And Current NRSP-7 Activity By Specie

†Approvals resulted in an additional 16 label claims for these aquatic species.

http://www.fda.gov/AnimalVeterinary/DevelopmentApprovalProcess/MinorUseMinorSpecies/ucm279396. htm

Source	80	Cash		In-Kind
Hatch Funding	\$	950,268		
NIFA/USDA	\$	858,000		
FDA/CVM(1)	\$	190,396	\$	1,625,000
Bionexus(2)			\$	370,000
CIDR Goat Efficacay(3)			\$	798,000
Pheasant Fenbendazole(4)			\$	850,000
Ivermectin Cattle Fever Tick(5)			\$	4,050,000
Merck (Feed Analyses)			\$	1,000
Alpharma (Feed Analyses)			\$	2,000
Pfizer (Feed Analyses & Ultrasour	nd)		\$	11,000
Tota	al = <b>"</b> \$	1,998,664	\$	7,707,000
Outside Cas	h = \$	1,048,396	1270	
Program Total (Cash + In Kind	) = \$	9,705,664		
Outside Cash)/Hatcl		1.10	Hatch Funds	= 48% Cash
Outside (Cash+In Kind)/Hatcl	h = 🗖	9.21	Hatch Fund	s = 10% total expenses

Table 3. Minor Use Animal Drug Program Total Funding 2009-2013 by Source

(1) Cash includes three MUMS grants and in-kind support includes 1.6 FTE assigned to the Program

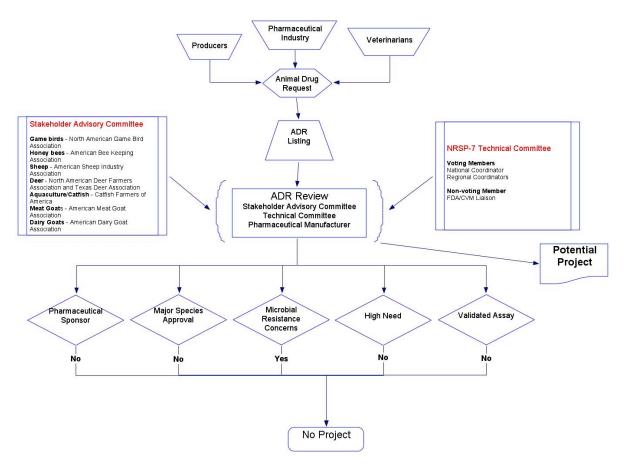
(2) National Coordinator's salary returned to the program and office overhead

(3) Produceres housing of goats duing study

(4) Cost to McFarland Farms for reproductive safety study

(5) Producers' cost for housing cattle

### Figure 1. Flow Chart Outlining The Process For Selection Of Drugs For Testing In The NRSP-7 Minor Use Animal Drug Program



#### Peer Reviewed Publications:

- 1. Rivera-Garcia, S., Angelos, J. A., Rowe, J. D., Byrne, B. A., Wetzlich, S. E., Van Liew, D. B., and Tell, L. A. (2014) Pharmacokinetics of ceftiofur crystalline-free acid following subcutaneous administration of a single dose to sheep, Am J Vet Res 75, 290-295.
- McDonnel, S. J., Tell, L. A., and Murphy, B. G. (2014) Pharmacokinetics and pharmacodynamics of suberoylanilide hydroxamic acid in cats, J Vet Pharmacol Ther 37, 196-200.
- Kinney, M. E., Lamberski, N., Wack, R., Foster, R., Neely, M., Tell, L., and Gehring, R. (2014) Population pharmacokinetics of a single intramuscular administration of tulathromycin in adult desert tortoises (Gopherus agassizii), J Vet Pharmacol Ther 37, 500-507.
- 4. Grismer, B., Rowe, J. D., Carlson, J., Wetzlich, S. E., and Tell, L. A. (2014) Pharmacokinetics of tulathromycin in plasma and milk samples after a single subcutaneous injection in lactating goats (Capra hircus), J Vet Pharmacol Ther 37, 205-208.
- Griffith, R., Yaeger, M., Hostetter, S., Tell, L. A., Wetzlich, S., Vickroy, T., Lillie, B., MacFarlane, W., Laudenslager, T., Whitley, E., Dzikamunhenga, R., and Larson, W. (2014) Safety of fenbendazole in Chinese ring-necked pheasants (Phasianus colchicus), Avian Dis 58, 8-15.
- 6. Wu, H., Baynes, R. E., Leavens, T., Tell, L. A., and Riviere, J. E. (2013) Use of population pharmacokinetic modeling and Monte Carlo simulation to capture individual animal variability in the prediction of flunixin withdrawal times in cattle, J Vet Pharmacol Ther 36, 248-257.
- Washburn, K. E., Fajt, V. R., Lawhon, S. D., Adams, L. G., Tell, L. A., and Bissett, W. T. (2013) Caprine abscess model of tulathromycin concentrations in interstitial fluid from tissue chambers inoculated with Corynebacterium pseudotuberculosis following subcutaneous or intrachamber administration, Antimicrobial agents and chemotherapy 57, 6295-6304.
- Spitsbergen, J. M., Frattini, S. A., Bowser, P. R., Getchell, R. G., Coffee, L. L., Wolfe, M. J., Fisher, J. P., Marinovic, S. J., and Harr, K. E. (2013) Epizootic neoplasia of the lateral line system of lake trout (Salvelinus namaycush) in New York's Finger Lakes, Vet Pathol 50, 418-433.
- 9. Snook, T. S., White, S. D., Hawkins, M. G., Tell, L. A., Wilson, L. S., Outerbridge, C. A., and Ihrke, P. J. (2013) Skin diseases in pet rabbits: a retrospective study of 334 cases seen at the University of California at Davis, USA (1984-2004), Veterinary dermatology.
- Shelver, W. L., Tell, L. A., Wagner, S., Wetzlich, S. E., Baynes, R. E., Riviere, J. E., and Smith, D. J. (2013) Comparison of ELISA and LC-MS/MS for the Measurement of Flunixin Plasma Concentrations in Beef Cattle after Intravenous and Subcutaneous Administration, J Agric Food Chem 61, 2679-2686.
- 11. McDonnel, S. J., Tell, L. A., and Murphy, B. G. (2013) Pharmacokinetics and pharmacodynamics of suberoylanilide hydroxamic acid in cats, J Vet Pharmacol Ther.
- Macpherson, M. L., Giguere, S., Hatzel, J. N., Pozor, M., Benson, S., Diaw, M., Sanchez, L. C., Vickroy, T. W., Tell, L., Wetzlich, S., and Sims, J. (2013) Disposition of desfuroylceftiofur acetamide in serum, placental tissue, fetal fluids, and fetal tissues after administration of ceftiofur crystalline free acid (CCFA) to pony mares with placentitis, J Vet Pharmacol Ther 36, 59-67.
- Grismer, B., Rowe, J. D., Carlson, J., Wetzlich, S. E., and Tell, L. A. (2013) Pharmacokinetics of tulathromycin in plasma and milk samples after a single subcutaneous injection in lactating goats (Capra hircus), J Vet Pharmacol Ther.
- Goetting, V., Lee, K. A., Woods, L., Clemons, K. V., Stevens, D. A., and Tell, L. A. (2013) Inflammatory marker profiles in an avian experimental model of aspergillosis, Medical mycology 51, 696-703.

- 15. DeDonder, K. D., Gehring, R., Baynes, R. E., Tell, L. A., Vickroy, T. W., Apley, M. D., and Riviere, J. E. (2013) Effects of new sampling protocols on procaine penicillin G withdrawal intervals for cattle, J Am Vet Med Assoc 243, 1408-1412.
- Dechant, J. E., Rowe, J. D., Byrne, B. A., Wetzlich, S. E., Kieu, H. T., and Tell, L. A. (2013) Pharmacokinetics of ceftiofur crystalline free acid after single and multiple subcutaneous administrations in healthy alpacas (Vicugna pacos), J Vet Pharmacol Ther 36, 122-129.
- 17. Cornwell, E. R., Labuda, S. L., Groocock, G. H., Getchell, R. G., and Bowser, P. R. (2013) Experimental Infection of Koi Carp with viral hemorrhagic septicemia virus type IVb, J Aquat Anim Health 25, 36-41.
- Cornwell, E. R., Bellmund, C. A., Groocock, G. H., Wong, P. T., Hambury, K. L., Getchell, R. G., and Bowser, P. R. (2013) Fin and gill biopsies are effective nonlethal samples for detection of viral hemorrhagic septicemia virus genotype IVb, J Vet Diagn Invest 25, 203-209.
- 19. Coffee, L. L., Casey, J. W., and Bowser, P. R. (2013) Pathology of tumors in fish associated with retroviruses: a review, Vet Pathol 50, 390-403.
- 20. Coffee, L. L., Bogdanovic, L. B., Cushing, T. L., and Bowser, P. R. (2013) Pharyngeal odontoma in an adult walleye (Sander vitreus), Vet Pathol 50, 483-487.
- 21. Topic Popovic, N., Howell, T., Babish, J. G., and Bowser, P. R. (2012) Cross-sectional study of hepatic CYP1A and CYP3A enzymes in hybrid striped bass, channel catfish and Nile tilapia following oxytetracycline treatment, Res Vet Sci 92, 283-291.
- Tell, L. A., Stephens, K., Teague, S. V., Pinkerton, K. E., and Raabe, O. G. (2012) Study of nebulization delivery of aerosolized fluorescent microspheres to the avian respiratory tract, Avian Dis 56, 381-386.
- 23. Romanet, J., Smith, G. W., Leavens, T. L., Baynes, R. E., Wetzlich, S. E., Riviere, J. E., and Tell, L. A. (2012) Pharmacokinetics and tissue elimination of tulathromycin following subcutaneous administration in meat goats, Am J Vet Res 73, 1634-1640.
- 24. Mehl, M. L., Tell, L., Kyles, A. E., Chen, Y. J., Craigmill, A., and Gregory, C. R. (2012) Pharmacokinetics and pharmacodynamics of A77 1726 and leflunomide in domestic cats, J Vet Pharmacol Ther 35, 139-146.
- 25. Lee, K. A., Tell, L. A., and Mohr, F. C. (2012) Inflammatory markers following acute fuel oil exposure or bacterial lipopolysaccharide in mallard ducks (Anas platyrhynchos), Avian Dis 56, 704-710.
- 26. Leclere, M., Magdesian, K. G., Cole, C. A., Szabo, N. J., Ruby, R. E., Rhodes, D. M., Edman, J., Vale, A., Wilson, W. D., and Tell, L. A. (2012) Pharmacokinetics and preliminary safety evaluation of azithromycin in adult horses, J Vet Pharmacol Ther 35, 541-549.
- Leavens, T. L., Tell, L. A., Clothier, K. A., Griffith, R. W., Baynes, R. E., and Riviere, J. E. (2012) Development of a physiologically based pharmacokinetic model to predict tulathromycin distribution in goats, J Vet Pharmacol Ther 35, 121-131.
- 28. Hope, K. L., Tell, L. A., Byrne, B. A., Murray, S., Wetzlich, S. E., Ware, L. H., Lynch, W., Padilla, L. R., and Boedeker, N. C. (2012) Pharmacokinetics of a single intramuscular injection of ceftiofur crystalline-free acid in American black ducks (Anas rubripes), Am J Vet Res 73, 620-627.
- 29. Dechant, J. E., Rowe, J. D., Byrne, B. A., Wetzlich, S. E., Kieu, H. T., and Tell, L. A. (2012) Pharmacokinetics of ceftiofur crystalline free acid after single and multiple subcutaneous administrations in healthy alpacas (Vicugna pacos), J Vet Pharmacol Ther.
- Cornwell, E. R., Anderson, G. B., Wooster, G. A., Getchell, R. G., Groocock, G. H., Casey, J. W., Bain, M. B., and Bowser, P. R. (2012) Low prevalence of Cyprinid Herpesvirus 3 Found in common carp (Cyprinus carpio carpio) collected from nine locations in the Great Lakes, J Wildl Dis 48, 1092-1096.
- 31. Clothier, K. A., Leavens, T., Griffith, R. W., Wetzlich, S. E., Baynes, R. E., Riviere, J. E., and Tell, L. A. (2012) Tulathromycin assay validation and tissue residues after single and multiple

subcutaneous injections in domestic goats (Capra aegagrus hircus), J Vet Pharmacol Ther 35, 113-120.

- 32. Bowser, P. R., Casey, J. W., Casey, R. N., Quackenbush, S. L., Lofton, L., Coll, J. A., and Cipriano, R. C. (2012) Swimbladder Leiomyosarcoma in Atlantic salmon (Salmo salar) in North America, J Wildl Dis 48, 795-798.
- 33. Young, G., Smith, G. W., Leavens, T. L., Wetzlich, S. E., Baynes, R. E., Mason, S. E., Riviere, J. E., and Tell, L. A. (2011) Pharmacokinetics of tulathromycin following subcutaneous administration in meat goats, Res Vet Sci 90, 477-479.
- 34. White, S. D., Bourdeau, P., Bruet, V., Kass, P. H., Tell, L., and Hawkins, M. G. (2011) Reptiles with dermatological lesions: a retrospective study of 301 cases at two university veterinary teaching hospitals (1992-2008), Veterinary dermatology 22, 150-161.
- 35. Tell, L. A., Brooks, J. W., Lintner, V., Matthews, T., and Kariyawasam, S. (2011) Antimicrobial susceptibility of Arcanobacterium pyogenes isolated from the lungs of whitetailed deer (Odocoileus virginianus) with pneumonia, J Vet Diagn Invest 23, 1009-1013.
- 36. Strunk, A., Imai, D. M., Osofsky, A., and Tell, L. A. (2011) Dysgerminoma in an eastern rosella (Platycercus eximius eximius), Avian Dis 55, 133-138.
- Leavens, T. L., Tell, L. A., Clothier, K. A., Griffith, R. W., Baynes, R. E., and Riviere, J. E. (2011) Development of a physiologically based pharmacokinetic model to predict tulathromycin distribution in goats, J Vet Pharmacol Ther.
- Kline, Y., Clemons, K. V., Woods, L., Stevens, D. A., and Tell, L. A. (2011) Pharmacokinetics of voriconazole in adult mallard ducks (Anas platyrhynchos), Medical mycology 49, 500-512.
- 39. Hawkins, M. G., Taylor, I. T., Byrne, B. A., Armstrong, R. D., and Tell, L. A. (2011) Pharmacokinetic-pharmacodynamic integration of orbifloxacin in Japanese quail (Coturnix japonica) following oral and intravenous administration, J Vet Pharmacol Ther 34, 350-358.
- 40. Goetting, V., Lee, K. A., and Tell, L. A. (2011) Pharmacokinetics of veterinary drugs in laying hens and residues in eggs: a review of the literature, J Vet Pharmacol Ther 34, 521-556.
- 41. Dore, E., Angelos, J. A., Rowe, J. D., Carlson, J. L., Wetzlich, S. E., Kieu, H. T., and Tell, L. A. (2011) Pharmacokinetics of ceftiofur crystalline free acid after single subcutaneous administration in lactating and nonlactating domestic goats (Capra aegagrus hircus), J Vet Pharmacol Ther 34, 25-30.
- 42. Cornwell, E. R., Eckerlin, G. E., Getchell, R. G., Groocock, G. H., Thompson, T. M., Batts, W. N., Casey, R. N., Kurath, G., Winton, J. R., Bowser, P. R., Bain, M. B., and Casey, J. W. (2011) Detection of viral hemorrhagic septicemia virus by quantitative reverse transcription polymerase chain reaction from two fish species at two sites in Lake Superior, J Aquat Anim Health 23, 207-217.
- Cornwell, E. R., Cinelli, M. J., McIntosh, D. M., Blank, G. S., Wooster, G. A., Groocock, G. H., Getchell, R. G., and Bowser, P. R. (2011) Epizootic Nocardia infection in cultured weakfish, Cynoscion regalis (Bloch and Schneider), J Fish Dis 34, 567-571.
- 44. Clothier, K. A., Leavens, T., Griffith, R. W., Wetzlich, S. E., Baynes, R. E., Riviere, J. E., and Tell, L. A. (2011) Pharmacokinetics of tulathromycin after single and multiple subcutaneous injections in domestic goats (Capra aegagrus hircus), J Vet Pharmacol Ther 34, 448-454.
- 45. Clothier, K. A., Leavens, T., Griffith, R. W., Wetzlich, S. E., Baynes, R. E., Riviere, J. E., and Tell, L. A. (2011) Tulathromycin assay validation and tissue residues after single and multiple subcutaneous injections in domestic goats (Capra aegagrus hircus), J Vet Pharmacol Ther.
- 46. Young, G., Smith, G. W., Leavens, T. L., Wetzlich, S. E., Baynes, R. E., Mason, S. E., Riviere, J. E., and Tell, L. A. (2010) Pharmacokinetics of tulathromycin following subcutaneous administration in meat goats, Res Vet Sci.
- Tell, L. A., Clemons, K. V., Kline, Y., Woods, L., Kass, P. H., Martinez, M., and Stevens, D. A. (2010) Efficacy of voriconazole in Japanese quail (Coturnix japonica) experimentally infected with Aspergillus fumigatus, Medical mycology 48, 234-244.

- 48. Rowe, J. D., Tell, L. A., Carlson, J. L., Griffith, R. W., Lee, K., Kieu, H., Wetzlich, S., and Hallford, D. (2010) Progesterone milk residues in goats treated with CIDR-G((R)) inserts, J Vet Pharmacol Ther 33, 605-609.
- Hope, K. M., Casey, R. N., Groocock, G. H., Getchell, R. G., Bowser, P. R., and Casey, J. W. (2010) Comparison of quantitative RT-PCR with cell culture to detect viral hemorrhagic septicemia virus (VHSV) IVb infections in the Great Lakes, J Aquat Anim Health 22, 50-61.
- 50. Clothier, K. A., Jordan, D. M., Loynachan, A. T., and Griffith, R. W. (2010) Safety evaluation of tulathromycin use in the caprine species: tulathromycin toxicity assessment in goats, J Vet Pharmacol Ther 33, 499-502.
- Bain, M. B., Cornwell, E. R., Hope, K. M., Eckerlin, G. E., Casey, R. N., Groocock, G. H., Getchell, R. G., Bowser, P. R., Winton, J. R., Batts, W. N., Cangelosi, A., and Casey, J. W. (2010) Distribution of an invasive aquatic pathogen (viral hemorrhagic septicemia virus) in the Great Lakes and its relationship to shipping, PLoS One 5, e10156.
- 52. Spitsbergen, J. M., Blazer, V. S., Bowser, P. R., Cheng, K. C., Cooper, K. R., Cooper, T. K., Frasca, S., Jr., Groman, D. B., Harper, C. M., Law, J. M., Marty, G. D., Smolowitz, R. M., St Leger, J., Wolf, D. C., and Wolf, J. C. (2009) Finfish and aquatic invertebrate pathology resources for now and the future, Comp Biochem Physiol C Toxicol Pharmacol 149, 249-257.
- 53. Rowe, J. D., Tell, L. A., and Wagner, D. C. (2009) Animal safety report on intravaginal progesterone controlled internal drug releasing devices in sheep and goats, J Vet Pharmacol Ther 32, 303-305.
- 54. Nguyen, K. Q., Hawkins, M. G., Taylor, I. T., Wiebe, V. J., and Tell, L. A. (2009) Stability and uniformity of extemporaneous preparations of voriconazole in two liquid suspension vehicles at two storage temperatures, Am J Vet Res 70, 908-914.
- 55. Kosoff, R. E., Chen, C. Y., Wooster, G. A., Getchell, R. G., Bowser, P. R., Clifford, A., Craig, J. L., Lim, P., Wetzlich, S. E., Craigmill, A. L., and Tell, L. A. (2009) Florfenicol residues in three species of fish after 10-day oral dosing in feed, J Aquat Anim Health 21, 8-13.
- 56. Davis, J. L., Smith, G. W., Baynes, R. E., Tell, L. A., Webb, A. I., and Riviere, J. E. (2009) Update on drugs prohibited from extralabel use in food animals, J Am Vet Med Assoc 235, 528-534.
- 57. Bowser, P. R., Kosoff, R. E., Chen, C. Y., Wooster, G. A., Getchell, R. G., Craig, J. L., Lim, P., Wetzlich, S. E., Craigmill, A. L., and Tell, L. A. (2009) Florfenicol residues in Nile tilapia after 10-d oral dosing in feed: effect of fish size, J Aquat Anim Health 21, 14-17.
- Wesley, I. V., Larsen, S., Hurd, H. S., McKean, J. D., Griffith, R., Rivera, F., Nannapaneni, R., Cox, M., Johnson, M., Wagner, D., and de Martino, M. (2008) Low prevalence of Listeria monocytogenes in cull sows and pork, J Food Prot 71, 545-549.
- 59. Smith, G. W., Davis, J. L., Tell, L. A., Webb, A. I., and Riviere, J. E. (2008) Extralabel use of nonsteroidal anti-inflammatory drugs in cattle, J Am Vet Med Assoc 232, 697-701.
- Ishak, H. D., Dumbacher, J. P., Anderson, N. L., Keane, J. J., Valkiunas, G., Haig, S. M., Tell, L. A., and Sehgal, R. N. (2008) Blood parasites in owls with conservation implications for the Spotted Owl (Strix occidentalis), PLoS One 3, e2304.
- H. S., Brudvig, J., Dickson, J., Mirceta, J., Polovinski, M., Matthews, N., and Griffith, R. (2008) Swine health impact on carcass contamination and human foodborne risk, Public Health Rep 123, 343-351.
- 62. Hawkins, M. G., Taylor, I. T., Craigmill, A. L., and Tell, L. A. (2008) Enantioselective pharmacokinetics of racemic carprofen in New Zealand white rabbits, J Vet Pharmacol Ther 31, 423-430.
- 63. Groocock, G. H., Grimmett, S. G., Getchell, R. G., Wooster, G. A., and Bowser, P. R. (2008) A survey to determine the presence and distribution of largemouth bass virus in wild freshwater bass in New York State, J Aquat Anim Health 20, 158-164.

- 64. Bartlett, S. L., Wooster, G. A., Sokolowski, M. S., Dove, A. D., and Bowser, P. R. (2008) Naturally occurring bacteraemia in American lobsters, Homarus americanus Milne-Edwards, in Long Island Sound, J Fish Dis 31, 19-25.
- 65. Needham, M. L., Webb, A. I., Baynes, R. E., Riviere, J. E., Craigmill, A. L., and Tell, L. A. (2007) Current update on drugs for game bird species, J Am Vet Med Assoc 231, 1506-1508.
- 66. Kosoff, R. E., Chen, C.-Y., Wooster, G. A., Getchell, R. G., Clifford, A., Craigmill, A. L., and P.R., B. (2007) Sulfadimethoxine and Ormetoprim Residues in Three Species of Fish After 5day Oral Dosing in Feed, J. Aquatic Animal Health, 109-115.
- 67. Getchell, R. G., Groocock, G. H., Schumacher, V. L., Grimmett, S. G., Wooster, G. A., and Bowser, P. R. (2007) Quantitative polymerase chain reaction assay for largemouth bass virus, J Aquat Anim Health 19, 226-233.
- 68. Chen, C.-Y., Chao, C.-B., and Bowser, P. R. (2007) Comparative histopathology of Streptococcus iniae and Streptococcus agalactiae-infected tilapia., Bull European Assoc Fish Pathologists 27, 2-9.
- 69. Sehgal, R. N., Hull, A. C., Anderson, N. L., Valkiunas, G., Markovets, M. J., Kawamura, S., and Tell, L. A. (2006) Evidence for cryptic speciation of Leucocytozoon spp. (Haemosporida, Leucocytozoidae) in diurnal raptors, The Journal of parasitology 92, 375-379.
- 70. Payne, M. A., Craigmill, A., Riviere, J. E., and Webb, A. I. (2006) Extralabel use of penicillin in food animals, J Am Vet Med Assoc 229, 1401-1403.
- 71. Hawkins, M. G., Kass, P. H., Zinkl, J. G., and Tell, L. A. (2006) Comparison of biochemical values in serum and plasma, fresh and frozen plasma, and hemolyzed samples from orange-winged Amazon parrots (Amazona amazonica), Veterinary clinical pathology / American Society for Veterinary Clinical Pathology 35, 219-225.
- 72. Chen, C.-Y., Chao, C.-B., and Bowser, P. R. (2006) Infection of tilapia Oreochromis sp. by Vibrio vulnificus in freshwater and low salinity environments. , J. World Aquaculture Soc 37, 82-88.
- 73. Wooster, G. A., Martinez, C. M., Ohara, D. S., and Bowser, P. R. (2005) Human Health Risks Associated with Formalin Treatments Used in Aquaculture: Initial Study North American J. Aquaculture 67, 111-113.
- 74. Tort, M. J., Hurley, D., Fernanzez-Cobas, C., Wooster, G. A., and Bowser, P. R. (2005) Effects of hydrogen peroxide treatment on catalase and glutathione activity in walleye (Sander vitreus). , J. World Aquaculture Soc 36, 577-586.
- 75. Tell, L. A., Craigmill, A. L., Clemons, K. V., Sun, Y., Laizure, S. C., Clifford, A., Ina, J. H., Nugent-Deal, J. P., Woods, L., and Stevens, D. A. (2005) Studies on itraconazole delivery and pharmacokinetics in mallard ducks (Anas platyrhynchos), J Vet Pharmacol Ther 28, 267-274.
- 76. Tell, L. A. (2005) Aspergillosis in mammals and birds: impact on veterinary medicine, Medical mycology 43 Suppl 1, S71-73.
- 77. Smith, G. W., Gehring, R., Craigmill, A. L., Webb, A. I., and Riviere, J. E. (2005) Extralabel intramammary use of drugs in dairy cattle, J Am Vet Med Assoc 226, 1994-1996.
- 78. Osofsky, A., Tell, L. A., Kass, P. H., Wetzlich, S. E., Nugent-Deal, J., and Craigmill, A. L. (2005) Investigation of Japanese quail (Coturnix japonica) as a pharmacokinetic model for cockatiels (Nymphicus hollandicus) and Poicephalus parrots via comparison of the pharmacokinetics of a single intravenous injection of oxytetracycline hydrochloride, J Vet Pharmacol Ther 28, 505-513.
- 79. KuKanich, B., Gehring, R., Webb, A. I., Craigmill, A. L., and Riviere, J. E. (2005) Effect of formulation and route of administration on tissue residues and withdrawal times, J Am Vet Med Assoc 227, 1574-1577.
- 80. Haskell, S. R., Payne, M., Webb, A., Riviere, J., and Craigmill, A. L. (2005) Antidotes in food animal practice, J Am Vet Med Assoc 226, 884-887.

- 81. Graham, J. E., Kollias-Baker, C., Craigmill, A. L., Thomasy, S. M., and Tell, L. A. (2005) Pharmacokinetics of ketoprofen in Japanese quail (Coturnix japonica), J Vet Pharmacol Ther 28, 399-402.
- Behring, R., van der Merwe, D., Pierce, A. N., Baynes, R. E., Craigmill, A. L., and Riviere, J. E. (2005) Multivariate meta-analysis of pharmacokinetic studies of ampicillin trihydrate in cattle, Am J Vet Res 66, 108-112.
- 83. Gehring, R., Haskell, S. R., Payne, M. A., Craigmill, A. L., Webb, A. I., and Riviere, J. E. (2005) Aminoglycoside residues in food of animal origin, J Am Vet Med Assoc 227, 63-66.
- 84. Chen, C.-Y., Wooster, G. A., Getchell, R. G., and Bowser, P. R. (2005) Distribution and depletion of oxytetracycline in two warm-water fish: tilapia and hybrid striped bass., J. World Aquaculture Soc 36, 564-569.
- 85. Chen, C.-Y., and Bowser, P. R. (2005) Pharmacokinetics of oxytetracycline in Nile tilapia (Oreochromis niloticus) challenged with Streptococcus iniae and Vibrio vulnificus., J. World Aquaculture Soc 36, 262-270.
- 86. Webb, A. I., Baynes, R. E., Craigmill, A. L., Riviere, J. E., and Haskell, S. R. (2004) Drugs approved for small ruminants, J Am Vet Med Assoc 224, 520-523.
- 87. Payne, M. A., Wetzlich, S. E., Robb, E. J., Brown, S. A., Gardner, I. A., Cullor, J. S., and Craigmill, A. L. (2004) Comparison of the use of regulatory assays and high-performance liquid chromatography for detection of residues of ceftiofur sodium metabolites in tissue specimens of culled dairy cattle, Am J Vet Res 65, 1730-1733.
- 88. Lane, V. M., Wetzlich, S., Clifford, A., Taylor, I., and Craigmill, A. L. (2004) Intravenous and subcutaneous pharmacokinetics of florfenicol in sheep, J Vet Pharmacol Ther 27, 191-196.
- 89. Lane, V. M., Villarroel, A., Wetzlich, S. E., Clifford, A., Taylor, I., and Craigmill, A. L. (2004) Intravenous and subcutaneous pharmacokinetics of florfenicol in sheep, J Vet Pharmacol Ther 27, 191-196.
- 90. Haskell, S. R., Payne, M. A., Webb, A. I., Riviere, J. E., and Craigmill, A. L. (2004) Current approved drugs for aquatic species, J Am Vet Med Assoc 224, 50-51.
- 91. Gehring, R., Baynes, R. E., Craigmill, A. L., and Riviere, J. E. (2004) Feasibility of using halflife multipliers to estimate extended withdrawal intervals following the extralabel use of drugs in food-producing animals, J Food Prot 67, 555-560.
- Drew, M. L., Waldrup, K., Kreeger, T., Craigmill, A. L., Wetzlich, S. E., and Mackintosh, C. (2004) Pharmacokinetics of ceftiofur in red deer (Cervus elaphus), J Vet Pharmacol Ther 27, 7-11.
- 93. Drew, M. L., Johnson, L., Pugh, D., Navarre, C. B., Taylor, I. T., and Craigmill, A. L. (2004) Pharmacokinetics of ceftiofur in llamas and alpacas, J Vet Pharmacol Ther 27, 13-20.
- 94. Dove, A. D., LoBue, C., Bowser, P., and Powell, M. (2004) Excretory calcinosis: a new fatal disease of wild American lobsters Homarus americanus, Dis Aquat Organ 58, 215-221.
- 95. Craigmill, A. L., Miller, G. R., Gehring, R., Pierce, A. N., and Riviere, J. E. (2004) Metaanalysis of pharmacokinetic data of veterinary drugs using the Food Animal Residue Avoidance Databank: oxytetracycline and procaine penicillin G, J Vet Pharmacol Ther 27, 343-353.
- 96. Chen, C.-Y., Wooster, G. A., and Bowser, P. R. (2004) Comparative blood chemistry and histopathology of tilapia infected with Vibrio vulnificus or Streptococcus iniae or exposed to carbon tetrachloride, gentamicin, or copper sulfate, Aquaculture 239, 421-443.
- 97. Chen, C.-Y., Getchell, R. G., Wooster, G. A., Craigmill, A. L., and Bowser, P. R. (2004) Oxytetracycline residues in four species of fish after 10-day oral dosing in feed, J. Aquatic Animal Health 16, 208-219.
- 98. Bowser, P. R., Wooster, G. A., Chen, C. Y., and Mo, R. S. (2004) Polymicrobic infection of hybrid striped bass (Morone chrysops x Morone saxatilis) with three bacterial pathogens: a case report, J Fish Dis 27, 123-127.

- 99. Bowser, P. R., Wooster, G. A., Chen, C.-Y., and Mo, R. S. (2004) Polymicrobic infection of hybrid striped bass (Morone chrysops X M. saxatilis) with three bacterial pathogens: A case study., J. Fish Diseases 27, 123-127.
- 100. Wang, J., Gehring, R., Baynes, R. E., Webb, A. I., Whitford, C., Payne, M. A., Fitzgerald, K., Craigmill, A. L., and Riviere, J. E. (2003) Evaluation of the advisory services provided by the Food Animal Residue Avoidance Databank, J Am Vet Med Assoc 223, 1596-1598.
- 101. Tort, M. J., Wooster, G. A., and Bowser, P. R. (2003) Effects of Hydrogen Peroxide on Hematology and Blood Chemistry Parameters of Walleye (Stizostedion vitreum), J. World Aquaculture Soc 34, 236-242.
- 102. Tort, M. J., Fletcher, C., Wooster, G. A., and Bowser, P. R. (2003) Stability of hydrogen peroxide in aquaria as a fish disease treatment, J. Applied Aquaculture 14, 37-45.
- 103. Tell, L. A., Woods, L., Foley, J., Needham, M. L., and Walker, R. L. (2003) A model of avian mycobacteriosis: clinical and histopathologic findings in Japanese quail (Coturnix coturnix japonica) intravenously inoculated with Mycobacterium avium, Avian Dis 47, 433-443.
- 104. Tell, L. A., Sun, Y., Needham, M., Johnson, J. R., and Shukla, A. (2003) In vivo release of oxytetracycline from a biodegradable controlled-release gel injected subcutaneously in Japanese quail (Coturnix coturnix japonica), J Vet Pharmacol Ther 26, 239-245.
- 105. Tell, L. A., Foley, J., Needham, M. L., and Walker, R. L. (2003) Diagnosis of avian mycobacteriosis: comparison of culture, acid-fast stains, and polymerase chain reaction for the identification of Mycobacterium avium in experimentally inoculated Japanese quail (Coturnix coturnix japonica), Avian Dis 47, 444-452.
- 106. Mehl, M. L., Kyles, A. E., Craigmill, A. L., Epstein, S., and Gregory, C. R. (2003) Disposition of cyclosporine after intravenous and multi-dose oral administration in cats, J Vet Pharmacol Ther 26, 349-354.
- 107. Kyles, A. E., Gregory, C. R., Craigmill, A. L., Griffey, S. M., Jackson, J., and Stanley, S. D. (2003) Pharmacokinetics of tacrolimus after multidose oral administration and efficacy in the prevention of allograft rejection in cats with renal transplants, Am J Vet Res 64, 926-934.
- 108. Hawkins, M. G., Wright, B. D., Pascoe, P. J., Kass, P. H., Maxwell, L. K., and Tell, L. A. (2003) Pharmacokinetics and anesthetic and cardiopulmonary effects of propofol in red-tailed hawks (Buteo jamaicensis) and great horned owls (Bubo virginianus), Am J Vet Res 64, 677-683.
- 109. Haskell, S. R., Gehring, R., Payne, M. A., Craigmill, A. L., Webb, A. I., Baynes, R. E., and Riviere, J. E. (2003) Update on FARAD food animal drug withholding recommendations, J Am Vet Med Assoc 223, 1277-1278.
- 110. Fielding, C. L., Magdesian, K. G., Elliott, D. A., Craigmill, A. L., Wilson, W. D., and Carlson, G. P. (2003) Pharmacokinetics and clinical utility of sodium bromide (NaBr) as an estimator of extracellular fluid volume in horses, J Vet Intern Med 17, 213-217.
- 111. Ferrell, S., Werner, J., Kyles, A., Lowenstine, L., Kass, P., and Tell, L. (2003) Evaluation of a collagen patch as a method of enhancing ventriculotomy healing in Japanese quail (Coturnix coturnix japonica), Veterinary surgery : VS 32, 103-112.
- 112. Downer, J., Craigmill, A., and Holstege, D. (2003) Toxic potential of oleander derived compost and vegetables grown with oleander soil amendments, Vet Hum Toxicol 45, 219-221.
- 113. Craigmill, A. L. (2003) A physiologically based pharmacokinetic model for oxytetracycline residues in sheep, J Vet Pharmacol Ther 26, 55-63.
- 114. Chen, C.-Y., Wooster, G. A., Getchell, R. G., Bowser, P. R., and Timmons, M. B. (2003) Blood Chemistry of Healthy, Nephrocalcinosis-affected, and Ozone-treated Tilapia in a Recirculation System, with Application of Discriminant Analysis, Aquaculture 218, 89-102.
- 115. Benson, K. G., Tell, L. A., Young, L. A., Wetzlich, S., and Craigmill, A. L. (2003) Pharmacokinetics of ceftiofur sodium after intramuscular or subcutaneous administration in green iguanas (Iguana iguana), Am J Vet Res 64, 1278-1282.

- 116. Tort, M. J., Pasnik, D., Fernandez-Cobas, C., Wooster, G. A., and Bowser, P. R. (2002) Quantitative scoring of gill pathology of walleyes (Stizostedion vitreum) exposed to hydrogen peroxide J. Aquatic Animal Health 14, 154-159.
- 117. Tort, M. J., Jennings-Bayshore, C., Wilson, D., Wooster, G. A., and Bowser, P., R. (2002) Assessing the effects of increasing hydrogen peroxide dosage on rainbow trout(Onchorhynchus mykiss) gills utilizing a digitized scoring methodology, J. Aquatic Animal Health 14, 95-103.
- 118. Shideler, S. E., Stoops, M. A., Gee, N. A., and Tell, L. A. (2002) Hematologic values for tule elk (Cervus elaphus nannodes), J Wildl Dis 38, 589-597.
- 119. Payne, M. A., Babish, J. G., Bulgin, M., Lane, M., Wetzlich, S., and Craigmill, A. L. (2002) Serum pharmacokinetics and tissue and milk residues of oxytetracycline in goats following a single intramuscular injection of a long-acting preparation and milk residues following a single subcutaneous injection, J Vet Pharmacol Ther 25, 25-32.
- 120. Martin-Jimenez, T., Baynes, R. E., Craigmill, A., and Riviere, J. E. (2002) Extrapolated withdrawal-interval estimator (EWE) algorithm: a quantitative approach to establishing extralabel withdrawal times, Regul Toxicol Pharmacol 36, 131-137.
- 121. Craigmill, A. L., and Cortright, K. A. (2002) Interspecies considerations in the evaluation of human food safety for veterinary drugs, AAPS PharmSci 4, E34.
- 122. Tripi, C. M., and Bowser, P., R. (2001) Toxicity of hydrogen peroxide to pre-exposed young-of-the-year walleye (Stizostedion vitreum): effects of water quality and age of fish, J. World Aquaculture Soc 32, 416-421.
- Paul, T. A., Burns, J. C., Shike, H., Getchell, R., Bowser, P. R., Whitlock, K. E., and Casey, J. W. (2001) Reporter gene expression in fish following cutaneous infection with pantropic retroviral vectors, Mar Biotechnol (NY) 3, S81-87.
- 124. Chen, C.-Y., Wooster, G. A., Getchell, R. G., and Bowser, P., R. (2001) Nephrocalcinosis in tilapia (Oreochromis niloticus) from a recirculation aquaculture system: a case report., J. Aquatic Animal Health 13, 368-372.
- 125. Aldana, L., Tsutsumi, V., Craigmill, A., Silveira, M. I., and Gonzalez de Mejia, E. (2001) alpha-Tocopherol modulates liver toxicity of the pyrethroid cypermethrin, Toxicol Lett 125, 107-116.
- 126. Saez, J. A., and Bowser, P. R. (2000) Hydrogen peroxide concentrations in hatchery culture units and effluent during and after treatment North American J. Aquaculture 63, 74-78.
- 127. Robbins, P. K., Tell, L. A., Needham, M. L., and Craigmill, A. L. (2000) Pharmacokinetics of piperacillan after intramuscular injection in red-tailed hawks (Buteo jamaicensis) and great horned owls (Bubo virginianus), J Zoo Wildl Med 31, 47-51.
- 128. Kyles, A. E., Gregory, C. R., and Craigmill, A. L. (2000) Comparison of the in vitro antiproliferative effects of five immunosuppressive drugs on lymphocytes in whole blood from cats, Am J Vet Res 61, 906-909.
- 129. Denver, M. C., Tell, L. A., Galey, F. D., Trupkiewicz, J. G., and Kass, P. H. (2000) Comparison of two heavy metal chelators for treatment of lead toxicosis in cockatiels, Am J Vet Res 61, 935-940.
- 130. Craigmill, A. L., Holland, R. E., Robinson, D., Wetzlich, S., and Arndt, T. (2000) Serum pharmacokinetics of oxytetracycline in sheep and calves and tissue residues in sheep following a single intramuscular injection of a long-acting preparation, J Vet Pharmacol Ther 23, 345-352.
- 131.Baynes, R. E., Payne, M., Martin-Jimenez, T., Abdullah, A. R., Anderson, K. L., Webb, A. I., Craigmill, A., and Riviere, J. E. (2000) Extralabel use of ivermectin and moxidectin in food animals, J Am Vet Med Assoc 217, 668-671.
- 132. Ringer, R. K., Miller, L. R., and Saylor, W. W. (1999) Minor-use animal drug program--a national agricultural program to approve animal drugs for minor species and uses, J Am Vet Med Assoc 214, 1636-1637.

- 133. Payne, M. A., Craigmill, A. L., Riviere, J. E., Baynes, R. E., Webb, A. I., and Sundlof, S. F. (1999) The Food Animal Residue Avoidance Databank (FARAD). Past, present and future, Vet Clin North Am Food Anim Pract 15, 75-88.
- 134. Payne, M. A., Baynes, R. E., Sundlof, S. E., CraigmillA, Webb, A. I., and Riviere, J. E. (1999) Drugs prohibited from extralabel use in food animals, J Am Vet Med Assoc 215, 28-32.
- 135. LaPierre, L. A., Holzschu, D. L., Bowser, P. R., and Casey, J. W. (1999) Sequence and transcriptional analyses of the fish retroviruses walleye epidermal hyperplasia virus types 1 and 2: evidence for a gene duplication, J Virol 73, 9393-9403.
- 136. Baynes, R. E., Martin-Jimenez, T., Craigmill, A. L., and Riviere, J. E. (1999) Estimating provisional acceptable residues for extralabel drug use in livestock, Regul Toxicol Pharmacol 29, 287-299.
- 137. Tort, M. J., Kuhl, A. J., Wooster, G. A., and Bowser, P. R. (1998) Modification of tolerance of walleye (Stizostedion vitreum) to bath treatment with hydrogen peroxide, J. World Aquaculture Soc 29, 499-504.
- 138. Tell, L., Harrenstien, L., Wetzlich, S., Needham, M., Nappier, J., Hoffman, G., Caputo, J., and Craigmill, A. (1998) Pharmacokinetics of ceftiofur sodium in exotic and domestic avian species, J Vet Pharmacol Ther 21, 85-91.
- 139. Smith, B. B. (1998) An overview of selected diseases and drug needs in the llama and alpaca industries, Vet Hum Toxicol 40 Suppl 1, 29-34.
- 140. Riviere, J. E., Webb, A. I., and Craigmill, A. L. (1998) Primer on estimating withdrawal times after extralabel drug use, J Am Vet Med Assoc 213, 966-968.
- 141. Rangel-Lugo, M., Payne, M., Webb, A. I., Riviere, J. E., and Craigmill, A. (1998) Prevention of antibiotic residues in veal calves fed colostrum, J Am Vet Med Assoc 213, 40-42.
- 142. Modric, S., Webb, A. I., and Derendorf, H. (1998) Pharmacokinetics and pharmacodynamics of tilmicosin in sheep and cattle, J Vet Pharmacol Ther 21, 444-452.
- 143. Miller, L. R., and Ringer, R. K. (1998) The NRSP-7 project as it relates to the aquaculture industry, Vet Hum Toxicol 40 Suppl 2, 2-3.
- 144.LaPierre, L. A., Holzschu, D. L., Wooster, G. A., Bowser, P. R., and Casey, J. W. (1998) Two closely related but distinct retroviruses are associated with walleye discrete epidermal hyperplasia, J Virol 72, 3484-3490.
- 145.Holzschu, D. L., Wooster, G. A., and Bowser, P. R. (1998) Experimental transmission of dermal sarcoma to the sauger Stizostedion canadense, Dis Aquat Organ 32, 9-14.
- 146. Fisher, J. P., Brown, S. B., Wooster, G. W., and Bowser, P. R. (1998) Maternal blood, egg and larval thiamin levels correlate with larval survival in landlocked Atlantic salmon, J Nutr 128, 2456-2466.
- 147. Aldana, L., Gonzalez de Mejia, E., Craigmill, A., Tsutsumi, V., Armendariz-Borunda, J., Panduro, A., and Rincon, A. R. (1998) Cypermethrin increases apo A-1 and apo B mRNA but not hyperlipidemia in rats, Toxicol Lett 95, 31-39.
- 148. (1998) Proceedings of the NRSP-7/FDA Workshop: Drug Availability for Minor Species in the 21st Century, Vet Hum Toxicol 40 Suppl 1, 1-63.
- 149. (1998) Proceedings of the 7th NRSP-7/USDA Workshop Drugs in Aquaculture: Current Status, Future Goals, Vet Hum Toxicol 40 Suppl 2, 1-64.
- 150. Stoffregen, D. A., Wooster, G. A., Bustos, P. S., Bowser, P. R., and Babish, J. G. (1997) Multiple route and dose pharmacokinetics of enrofloxacin in juvenile Atlantic salmon, J Vet Pharmacol Ther 20, 111-123.
- 151. Riviere, J. E., Martin-Jimenez, T., Sundlof, S. F., and Craigmill, A. L. (1997) Interspecies allometric analysis of the comparative pharmacokinetics of 44 drugs across veterinary and laboratory animal species, J Vet Pharmacol Ther 20, 453-463.
- 152. Quackenbush, S. L., Holzschu, D. L., Bowser, P. R., and Casey, J. W. (1997) Transcriptional analysis of walleye dermal sarcoma virus (WDSV), Virology 237, 107-112.

- 153. Martin-Jimenez, T., Craigmill, A. L., and Riviere, J. E. (1997) Extralabel use of oxytetracycline, J Am Vet Med Assoc 211, 42-44.
- 154. Holzschu, D. L., Fodor, S. K., Quackenbush, S. L., Earnest-Koons, K., Bowser, P. R., Vogt, V. M., and Casey, J. W. (1997) Molecular characterization of a piscine retrovirus, walleye dermal sarcoma virus, Leukemia 11 Suppl 3, 172-175.
- 155. Earnest-Koons, K. A., Schachte, J. H., Jr., and Bowser, P. R. (1997) Lymphosarcoma in a brook trout, J Wildl Dis 33, 666-669.
- 156. Damian, P., Craigmill, A. L., and Riviere, J. E. (1997) Extralabel use of nonsteroidal antiinflammatory drugs, J Am Vet Med Assoc 211, 860-861.
- 157. Craigmill, A. L., Rangel-Lugo, M., Damian, P., and Riviere, J. E. (1997) Extralabel use of tranquilizers and general anesthetics, J Am Vet Med Assoc 211, 302-304.
- 158. Craigmill, A. L., Brown, S. A., Wetzlich, S. E., Gustafson, C. R., and Arndt, T. S. (1997) Pharmacokinetics of ceftiofur and metabolites after single intravenous and intramuscular administration and multiple intramuscular administrations of ceftiofur sodium to sheep, J Vet Pharmacol Ther 20, 139-144.
- 159. Courtin, F., Craigmill, A. L., Wetzlich, S. E., Gustafson, C. R., and Arndt, T. S. (1997) Pharmacokinetics of ceftiofur and metabolites after single intravenous and intramuscular administration and multiple intramuscular administrations of ceftiofur sodium to dairy goats, J Vet Pharmacol Ther 20, 368-373.
- 160.Baynes, R. E., Craigmill, A. L., and Riviere, J. E. (1997) Residue avoidance after topical application of veterinary drugs and parasiticides, J Am Vet Med Assoc 210, 1288-1289.
- 161. Vancutsem, P. M., and Babish, J. G. (1996) In vitro and in vivo study of the effects of enrofloxacin on hepatic cytochrome P-450. Potential for drug interactions, Vet Hum Toxicol 38, 254-259.
- 162. Sweeney, L. M., Shuler, M. L., Quick, D. J., and Babish, J. G. (1996) A preliminary physiologically based pharmacokinetic model for naphthalene and naphthalene oxide in mice and rats, Ann Biomed Eng 24, 305-320.
- 163. Stoffregen, D. A., Bowser, P. R., and Babish, J. G. (1996) Antibacterial chemotherapeutants for finfish aquaculture: a synopsis of laboratory and field efficacy and safety studies, J. Aquatic Animal Health 8, 181-207.
- 164. Stoffregen, D. A., Bachman, S. C., Perhman, R. E., Bowser, P. R., and Babish, J. G. (1996) Initial disease report of Streptococcus iniae infection in hybrid striped (sunshine) bass and successful therapeutic intervention with the fluoroquinolone antibacterial enrofloxacin, J. World Aquaculture Soc 27, 420-434.
- 165. Poulet, F. M., Bowser, P. R., and Casey, J. W. (1996) PCR and RT-PCR analysis of infection and transcriptional activity of walleye dermal sarcoma virus (WDSV) in organs of adult walleyes (Stizostedion vitreum), Vet Pathol 33, 66-73.
- 166. Gonzalez de Mejia, E., and Craigmill, A. L. (1996) Transfer of lead from lead-glazed ceramics to food, Arch Environ Contam Toxicol 31, 581-584.
- 167.Zelikoff, J. T., Bowser, D., Squibb, K. S., and Frenkel, K. (1995) Immunotoxicity of low level cadmium exposure in fish: an alternative animal model for immunotoxicological studies, J Toxicol Environ Health 45, 235-248.
- 168. Trepanier, L. A., and Babish, J. G. (1995) Pharmacokinetic properties of bromide in dogs after the intravenous and oral administration of single doses, Res Vet Sci 58, 248-251.
- 169. Trepanier, L. A., and Babish, J. G. (1995) Effect of dietary chloride content on the elimination of bromide by dogs, Res Vet Sci 58, 252-255.
- 170. Poulet, F. M., Vogt, V. M., Bowser, P. R., and Casey, J. W. (1995) In situ hybridization and immunohistochemical study of walleye dermal sarcoma virus (WDSV) nucleic acids and proteins in spontaneous sarcomas of adult walleyes (Stizostedion vitreum), Vet Pathol 32, 162-172.

- 171. Marley, S. E., Knapp, S. E., Rognlie, M. C., Thompson, J. R., Stoppa, T. M., Button, S. M., Wetzlich, S., Arndt, T., and Craigmill, A. (1995) Efficacy of ivermectin pour-on against Ostertagia ostertagi infection and residues in the American bison, Bison bison, J Wildl Dis 31, 62-65.
- 172. Hsu, H.-M., Bowser, P. R., Schachte, J., J.H., Scarlett, J. M., and Babish, J. G. (1995) Winter field trials of enrofloxacin for the control of Aeromonas salmonicida infection in salmonids, J. World Aquaculture Soc 26, 307-314.
- 173. Holzschu, D. L., Martineau, D., Fodor, S. K., Vogt, V. M., Bowser, P. R., and Casey, J. W. (1995) Nucleotide sequence and protein analysis of a complex piscine retrovirus, walleye dermal sarcoma virus, J Virol 69, 5320-5331.
- 174. Rumsey, G. L., Siwicki, A. K., Anderson, D. P., and Bowser, P. R. (1994) Effect of soybean protein on serological response, non-specific defense mechanisms, growth, and protein utilization in rainbow trout, Vet Immunol Immunopathol 41, 323-339.
- 175. Hsu, H.-M., Wooster, G. A., and Bowser, P. R. (1994) Efficacy of enrofloxacin in salmonids for treatment of bacterial kidney disease caused by Renibacterium salmoninarum, J. Aquatic Animal Health. 6, 220-223.
- 176. Hsu, H.-M., Wooster, G. A., and Bowser, P., R. (1994) Tissue distribution of enrofloxacin in fingerling rainbow trout (Oncorhynchus mykiss) following different doses of oral administration, J. World Aquacultural Society 25, 535-540.
- 177.Bowser, P. R., Wooster, G. A., and Hsu, H.-M. (1994) Laboratory efficacy of enrofloxacin for the control of Aeromonas salmonicida infection in rainbow trout (Oncorhynchus mykiss), J. Aquatic Animal Health 6, 288-291.
- 178. Bowser, D. H., Frenkel, K., and Zelikoff, J. T. (1994) Effects of in vitro nickel exposure on the macrophage-mediated immune functions of rainbow trout (Oncorhynchus mykiss), Bull Environ Contam Toxicol 52, 367-373.
- 179. Vancutsem, P. M., and Babish, J. G. (1993) Effects of ciprofloxacin and enrofloxacin on zoxazolamine kinetics, plasma concentration and sleeping times in mice, Toxicol Lett 69, 1-14.
- 180. Hsu, H.-M., Stroffregen, D. A., Wooster, G. A., and Bowser, P., R. (1993) Target animal safety studies of enrofloxacin in fingerling rainbow trout (Oncorhynchus mykiss), Bull European Assoc Fish Pathologists 13, 141-143.
- 181. Eltom, S. E., Babish, J. G., and Schwark, W. S. (1993) The postnatal development of drugmetabolizing enzymes in hepatic, pulmonary and renal tissues of the goat, J Vet Pharmacol Ther 16, 152-163.
- 182. Martineau, D., Bowser, P. R., Renshaw, R. R., and Casey, J. W. (1992) Molecular characterization of a unique retrovirus associated with a fish tumor, J Virol 66, 596-599.
- 183. Craigmill, A. L., Pass, M. A., and Wetzlich, S. (1992) Comparative pharmacokinetics of amoxicillin administered intravenously to sheep and goats, J Vet Pharmacol Ther 15, 72-77.
- 184. Bowser, P. R., Wooster, G. A., St Leger, J., and Babish, J. G. (1992) Pharmacokinetics of enrofloxacin in fingerling rainbow trout (Oncorhynchus mykiss), J Vet Pharmacol Ther 15, 62-71.
- 185. Zelikoff, J. T., Enane, N. A., Bowser, D., Squibb, K. S., and Frenkel, K. (1991) Development of fish peritoneal macrophages as a model for higher vertebrates in immunotoxicological studies. I. Characterization of trout macrophage morphological, functional, and biochemical properties, Fundam Appl Toxicol 16, 576-589.
- 186. Sundlof, S. F., Craigmill, A. L., and Riviere, J. E. (1991) Use of the food animal residue avoidance databank, J Am Vet Med Assoc 198, 816-819.
- 187. Galey, F. D., Tracy, M. L., Craigmill, A. L., Barr, B. C., Markegard, G., Peterson, R., and O'Connor, M. (1991) Staggers induced by consumption of perennial ryegrass in cattle and sheep from northern California, J Am Vet Med Assoc 199, 466-470.

- 188. Bulgin, M. S., Lane, V. M., Archer, T. E., Baggot, J. D., and Craigmill, A. L. (1991) Pharmacokinetics, safety and tissue residues of sustained-release sulfamethazine in sheep, J Vet Pharmacol Ther 14, 36-45.
- 189.\Bowser, P. R., and Babish, J. G. (1991) Enrofloxacin in salmonids, Vet Hum Toxicol 33 Suppl 1, 46-48.
- 190. Vancutsem, P. M., Babish, J. G., and Schwark, W. S. (1990) The fluoroquinolone antimicrobials: structure, antimicrobial activity, pharmacokinetics, clinical use in domestic animals and toxicity, Cornell Vet 80, 173-186.
- 191. Bowser, P. R., Schachte, J., J.H., Wooster, G. A., and Babish, J. G. (1990) Experimental treatment of Aeromonas salmonicida infections with enrofloxacin and oxolinic acid: field trials, J. Aquat. Animal Health 2, 198-203.
- 192. Bowser, P. R., and House, M. (1990) In vitro sensitivity of some fish pathogens to the quinolones, nalidixic acid and oxolinic acid and the fluoroquinolone enrofloxacin. , Bull. Europ. Assoc. Fish Pathol. 10, 48-49.
- 193. Bowser, P. R., and Babish, J. G. (1990) Clinical pharmacology and efficacy of fluoroquinolones in fish, Ann. Rev. Fish Diseases 1, 63-66.
- 194. Babish, J. G., Coles, G. C., Tritschler, J. P., 2nd, Gutenmann, W. H., and Lisk, D. J. (1990) Toxicity and tissue residue depletion of levamisole hydrochloride in young goats, Am J Vet Res 51, 1126-1130.
- 195. Trotter, E. J., Crissman, J., Robson, D., and Babish, J. (1988) Influence of nonbiologic implants on laminectomy membrane formation in dogs, Am J Vet Res 49, 634-643.
- 196. Bowser, P. R., Landy, R. B., Wooster, G. A., and Babish, J. G. (1988) Efficacy of elevated dietary fluoride for the control of Renibacterium salmoninarum infection in rainbow trout Salmo gairdneri., J. World Aquacult. Soc. 19, 1-7.
- 197. Shoaf, S. E., Schwark, W. S., Guard, C. L., and Babish, J. G. (1987) The development of hepatic drug-metabolizing enzyme activity in the neonatal calf and its effect on drug disposition, Drug Metab Dispos 15, 676-681.
- 198. Conzelman, G. M., Babish, J. G., Davidson, J. N., McMillan, R. A., and Copeland, D. D. (1987) Pharmacokinetics of 1-cyclopropyl-6-fluoro-1,4-dihydro-4-oxo-7-(4-ethyl-1-piperazinyl)-3-q uinoline- carboxylic acid (BAY Vp 2674) in chickens, Proc West Pharmacol Soc 30, 393-395.
- 199. Sundlof, S. F., Riviere, J. E., Craigmill, A. L., and Buck, W. B. (1986) Computerized foodanimal residue-avoidance data bank for veterinarians, J Am Vet Med Assoc 188, 73-76.
- 200. Sundlof, S. F., Craigmill, A. C., and Riviere, J. E. (1986) Food Animal Residue Avoidance Databank (FARAD): a pharmacokinetic-based information resource, J Vet Pharmacol Ther 9, 237-245.
- 201. Stoewsand, G. S., Babish, J. G., Telford, J. N., Bahm, C., Bache, C. A., Gutenmann, W. H., and Lisk, D. J. (1986) Response of Japanese quail fed seed meal from sunflowers grown on a municipal sludge-amended soil: elevation of cadmium in tissues, J Toxicol Environ Health 17, 91-100.
- 202. McLain, D. E., Babish, J. G., and Roe, D. A. (1985) Pharmacokinetics of ethanol in the ferret, Alcohol Clin Exp Res 9, 138-142.
- 203. Kemen, M. J., Frank, R. A., and Babish, J. B. (1985) An outbreak of equine influenza at a harness horse racetrack, Cornell Vet 75, 277-288.
- 204. Frederick, K. A., and Babish, J. G. (1985) Compendium of recent literature on the ferret, Lab Anim Sci 35, 298-318.
- 205. Patton, W. H., Schwartz, L. D., Babish, J. G., and Lisk, D. J. (1984) Use of amprolium for the control of coccidiosis in pheasants, Avian Dis 28, 693-699.
- 206. Cottrell, W. O., Ringer, R. K., and Babish, J. G. (1984) Acute toxicity of dietary polybrominated biphenyls in Bobwhite quail, Bull Environ Contam Toxicol 33, 308-312.

207. Burrows, G. E., Tyler, R. D., Craigmill, A. L., and Barto, P. B. (1984) Chloramphenicol and the neonatal calf, Am J Vet Res 45, 1586-1591.

208. Wilson, R. P., Bowser, P. R., and Poe, W. E. (1983) Dietary pantothenic acid requirement of fingerling channel catfish, J Nutr 113, 2124-2128.

- 209. Skrabalak, D. S., and Babish, J. G. (1983) Safety standards for occupational exposure to dichloromethane, Regul Toxicol Pharmacol 3, 139-143.
- 210. Davidson, J. N., Babish, J. G., and Dunny, G. M. (1982) Bovine mastitis: antimicrobial resistance patterns, J Am Vet Med Assoc 180, 153-155.
- 211. Davidson, J. N., and Babish, J. G. (1982) Clinical use of odds ratios in selecting antimicrobial therapy for bovine Pasteurella pneumonia, Am J Vet Res 43, 922-923.

#### Literature Cited

1. Ringer, R. K., Miller, L. R., and Saylor, W. W. (1999) Minor-use animal drug program--a national agricultural program to approve animal drugs for minor species and uses, J Am Vet Med Assoc 214, 1636-1637.

2. Lathers, C. M. (2003) Challenges and opportunities in animal drug development: a regulatory perspective, Nature reviews. Drug discovery 2, 915-918.

3. Pfizer. Personal Communication (2013).

4. AHI. Animal Health Institue Statistics.

### External Links

NRSP-7: http://www.nrsp-7.org

FARAD: http://www.farad.org

CDC information on food borne antibiotic resistance [http://www.cdc.gov/narms/animals.html]

FDA information on antibiotic use in food animals .[http://www.fda.gov/AnimalVeterinary/%20SafetyHealth/AntimicrobialResistance/JudiciousUseo fAntimicrobials/default.htm]

American Veterinary Medical Association position on use ofantibiotics in food producing animals. [https://www.avma.org/KB/Policies/Pages/Judicious-Therapeutic-Use-of-Antimicrobials.aspx']

Public Master Files supporting approved New Animal Drug Applications for Minor Uses http://www.fda.gov/AnimalVeterinary/DevelopmentApprovalProcess/MinorUseMinorSpecies/ucm279396.htm

Public Master Files (PMFs) in development http://www.fda.gov/AnimalVeterinary/DevelopmentApprovalProcess/MinorUseMinorSpecies/ucm287667.htm

Animal Health Institute Statistics: http://www.ahi.org/about-animal-medicines/industry-statistics/

Cattle Fever Tick Information http://www.ars.usda.gov/is/AR/archive/nov10/ticks1110.htm

https://www.federalregister.gov/articles/2013/07/24/2013-17804/environmental-impact-statement-proposed-cattle-fever-tick-control-barrier-in-south-texas

Honey Bees

http://www.nrdc.org/wildlife/animals/bees.asp?gclid=Cly\_ieDX-7sCFeg-MgodpTAAYw

Global Veterinary Health Products Market to Spike to Over \$28 Billion by 2017 http://finance.yahoo.com/news/global-veterinary-health-products-market-193000008.html Appendix E

NRSP-7: A National Agricultural Program for Minor Use Animal Drugs

Administrative Advisors: Margaret E. Smith (Chair), North Eastern; Philip Elzer, Southern; Frank D. Galey, Western: George Smith, North Central.

				Rese	arch									
								_				roje		
		CI	RIS Coo	des	P	ersonn	el	Ex	tension		Obj	ecti	ves	
Participant Name	Institution and								National	1	2	3	4	5
[email address]	Department	RPA	SOI	FOS	SY	PY	ΤY	FTE	Program					
Lisa Tell	UC Davis, College of Veterinary	711	3099	1180										
latell@ucdavis.edu	Medicine, Dept Medicine &	711	3820	1180										
	Epidemiology	711	3799	1180										
		711	3699	1180	0.1	0.0	1.0	0.0		1	2	3	4	
		711	3299	1180										
		711	3099	1180										
Thomas W. Vickroy	U. Florida, College of Veterinary	711	3820	1180										
vickroy@ufl.edu	Medicine, Dept Physiological	711	3699	1180										
	Sciences.	311	3910	1180	0.15	0.15	0.15	0.05		1	2	3	4	
		308	3099	1180										
		711	3899	1180										
Ronald W. Griffith	Iowa State U, College of	311	3910	1180	0.3	0.5	0.0	0.0		1	2	3	4	
rgriffit@iastate.edu	Veterinary Medicine, Dept Vet	301	3820	1100										
	Micro & Preventive Medicine													
Rod Getchell	Cornell University College of	311	3910	1060	0.1	0.0	0.0	0.0		1	2	3	4	
rgg4@cornell.edu	Veterinary Medicine, Dept													
	Microbiology & Immunology													

#### Appendix F: NRSP-7/MINOR USE ANIMAL DRUG PROGRAM BUDGET REQUESTS SUMMARY

(01 October 1, 2015 – September 30, 2020)

## NRSP – 7 Minor Use Animal Drugs

A National Agricultural Program for Minor Use Animal Drugs

				MRF F	UNDING						
DESCRIPTION	RIPTION Proposed FY 1 Proposed FY 2 FY 2015-2016 FY 2015-2016				Propose FY 201		Propose FY 201		Proposed FY 5 FY 2019-2020		
	Dollars	FTE	Dollars	FTE	Dollars	FTE	Dollars	FTE	Dollars	FTE	
SALARIES <sup>(a)</sup>	76,900	2.5	76,900	2.5	76,900	2.5	76,900	2.5	76,900	2.5	
FRINGE											
BENEFITS	24,768		24,768		24,768		24,768		24,768		
WAGES											
TRAVEL <sup>(b)</sup>	5,000		5,000		5,000		5,000		5,000		
RESEARCH											
	213,841		213,841		213,841		213,841		213,841		
MAINTENANCE <sup>(d)</sup>	3,096		3,096		3,096		3,096		3,096		
EQUIPMENT/											
CAPITAL											
	1,395		1,395		1,395		1,395		1,395		
TOTAL	325,000	2.5	325,000	2.5	325,000	2.5	325,000	2.5	325,000	2.5	

<sup>(a)</sup>Includes part-time salaries and support staff in Northeast, Southern and Western Regions. <sup>(b)</sup>Travel for GLP monitoring of studies and final reports. <sup>(c)</sup>Funding of Target Animal Safety, Human Food Safety and Residue Depletion studies.

<sup>(d)</sup>Maintenance contracts for analytical equipment.

<sup>(e)</sup>Leasing of analytical equipment.

Please check	one of the fol	US F	□ Industry	∎ Fec g Administ	tration/Cente	es FDA/CV	M		ts 🗆 SAI	ESs
DESCRIPTION	Propose FY 2015		Propose FY 2015		Propose FY 2017		Propose FY 2018		Propose FY 2019	
	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>
SALARIES AND WAGES	222,720	1.6	222,720	1.6	222,720	1.6	222,720	1.6	222,720	1.6
FRINGE BENEFITS	88,688		88,688		88,688		88,688		88,688	
TRAVEL	5,000		5,000		5,000		5,000		5,000	
MATERIALS AND SUPPLIES	9,000		9,000		9,000		9,000		9,000	
PUBLICATIONS										
CAPITAL EQUIPMENT										
OTHER DIRECT COSTS										
RESEARCH										
GOV'T HOLD BACK										
TOTAL	325,408	1.6	325,408	1.6	325,408	1.6	325,408	1.6	325,408	1.6

(a) Salary, benefits, materials and supplies for full-time FDA/CVM liaison and assistant to the NRSP-7 program provided by FDA/CVM.

Please c	heck one of th	-		ustry [	CES OF FUN I Federal Ag ege and Stat	gencies [	□ Grants/C	Contracts	SAESs	
DESCRIPTION	Propose	d FY 1	Propose	ed FY 2	Propose	ed FY 3	Propos	ed FY 4	Propose	ed FY 5
	FY 2015		FY 201	5-2016	FY 2017	7-2018	FY 201	8-2019	FY 2019	9-2020
	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>	Dollars	Dollars	FTE <sup>(a)</sup>	Dollars	FTE <sup>(a)</sup>	Dollars
SALARIES AND WAGES	53,840	0.4	53,840	0.4	53,840	0.4	53,840	0.4	53,840	0.4
FRINGE BENEFITS	35,360		35,360		35,360		35,360		35,360	
TRAVEL										
MATERIALS AND SUPPLIES										
PUBLICATIONS										
CAPITAL EQUIPMENT										
OTHER DIRECT COSTS										
RESEARCH										
GOV'T HOLD BACK										
TOTAL	89,200	0.4	89,200	0.4	89,200	0.4	89,200	0.4	89,200	0.4

(a) Salary and benefits for four regional coordinators to the NRSP-7 program provided by individual college and state funding.

Please ch	eck one of the	e following			RCES OF FUNE □ Federal Aç		□ Grants/C	Contracts	□ SAESs			
			ther (please lis	st):								
DESCRIPTION	Proposed FY 1 FY 2015-2016			Proposed FY 2 FY 2015-2016		Proposed FY 3 FY 2017-2018				ed FY 4 3-2019	Propose FY 2019	
-	Dollars	FTE	Dollars	FTE	Dollars	FTE	Dollars	FTE	Dollars	FTE		
SALARIES AND WAGES <sup>(a)</sup>	102,990	0.5	102,990	0.5	102,990	0.5	102,990	0.5	102,990	0.5		
FRINGE BENEFITS	29,867		29,867		29,867		29,867		29,867			
TRAVEL												
RESEARH MATERIALS AND SUPPLIES	982,800 <sup>(b)</sup>		982,800 <sup>(b)</sup>		982,800 <sup>(b)</sup>		982,800 <sup>(b)</sup>		982,800 <sup>(b)</sup>			
PUBLICATIONS												
CAPITAL EQUIPMENT												
OTHER DIRECT COSTS												
SUPPORT R&D <sup>(c)</sup>	(C)		(c)		(C)		(c)		(C)			
TOTAL	1,115,657	0.5	1,115,657	0.5	1,115,657	0.5	1,115,657	0.5	1,115,657	0.5		

(a) Includes personnel from the veterinary pharmaceutical and animal production industries needed to review protocols, data submissions, change label claims and file amended anima drug applications. Companies specifically involved with NRSP-7 include Pfizer, Biomedia, Intervet, Schering and Alpharma.

(b) For the years 2009 to 2014, the average **MATERIALS AND SUPPLIES** contribution from industry has increased to \$982,800/year. **MATERIALS AND SUPPLIES** included total costs directly attributable to carrying out the grant including storage and office space rental, supplying drugs, analytical support or animals for efficacy, safety or residue depletion studies.

(c) Cooperation with pharmaceutical companies to sponsor animal drug research projects is vital to the NRSP-7 program. The major contribution to the program is the cost borne by the pharmaceutical industry for the approval of drugs for a major species, estimated at approximately \$100 million or more per approved drug.